



(RESEARCH ARTICLE)



DESIGN, STRUCTURAL ELUCIDATION, AND TARGETED CYTOTOXIC EVALUATION OF NOVEL METAL–SCHIFF BASE COMPLEXES AS POTENTIAL ANTICANCER AGENTS

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ABSTRACT

Novel Fe(III), Mn(II), Co(II), Ni(II), Cu(II), Cd(II), Mg(II), Al(III), Zn(II), and Ba(II) complexes of an amido-salicylaldehyde ligand were synthesized and characterized by various analytical and spectroscopic techniques. The ligand coordinated as a neutral or dibasic pentadentate donor through imino nitrogen, hydroxyl oxygen, and amide oxygen atoms, forming distorted octahedral complexes. Conductivity measurements indicated non-electrolytic behavior. ESR studies revealed axial Cu(II) spectra with a dx^2-y^2 ground state, while Fe(III), Co(II), and Mn(II) complexes showed isotropic signals. Cytotoxicity studies against MCF-7 cells demonstrated enhanced activity after complexation, with the Zn(II) complex showing the highest potency compared with vinblastine sulfate. Molecular docking further supported the experimental cytotoxicity results through favorable binding interactions.

Keywords: Amido-Salicylaldehyde, Complexes, Spectra, ESR, Magnetism, Conductivity, Cytotoxicity, Molecular Docking.

1 INTRODUCTION

Following the breakthrough discovery and clinical application of cisplatin as a mainstay chemotherapeutic agent, extensive research has been directed toward the development of non-platinum-based metallo drugs utilizing transition metals such as gold, ruthenium, copper, and zinc to overcome the severe systemic toxicities, off-target side effects, and

intrinsic or acquired resistance associated with platinum therapies [1]. These coordination compounds exert potent antitumor effects through diverse mechanisms, including modulating cellular metal ion homeostasis, depleting intracellular trace elements, and perturbing key signaling pathways governing malignant cell proliferation [2]. In this context, hydrazone ligands and their coordination complexes have emerged as highly versatile scaffolds in medicinal chemistry due to their facile synthesis, superior hydrolytic stability compared to conventional imines, and rich coordination chemistry provided by polydentate N,O-donor atom sets [3-5]. Moreover, complexation with transition metals significantly amplifies the intrinsic biological efficacy of parent hydrazone moieties [6]. Given that breast cancer (BC) remains the second most prevalent neoplasm globally and the leading cause of cancer-related mortality among women exhibiting distinct epidemiological profiles with lower median ages at diagnosis across Arab and African demographics, including Egypt there is an urgent imperative for novel targeted therapeutics [7-9]. Addressing this need, the present study details the design, synthesis, and comprehensive spectroscopic characterization of a novel series of transition metal complexes derived from an alanine-based hydrazone ligand. Furthermore, the cytotoxic potential of these synthesized complexes was rigorously evaluated against human breast adenocarcinoma (MCF-7) cell lines, complemented by *in silico* molecular docking simulations to elucidate their binding interactions and plausible mechanisms of action within key biological targets.

2 Experimental

2.1 Ligand synthesis

The ligand (**HL**) was synthesized through a three-step procedure. First, α -alanine was esterified in ethanol using concentrated H_2SO_4 under reflux for 3 h. The resulting α -alanine ester was then reacted with hydrazine under reflux at **70 °C for 3 h** to form the corresponding hydrazide. Finally, the product was reacted with salicylaldehyde in ethanol under reflux at **70 °C for 2–3 h**. After cooling, the precipitated product was filtered and dried to obtain the desired **HL ligand**.

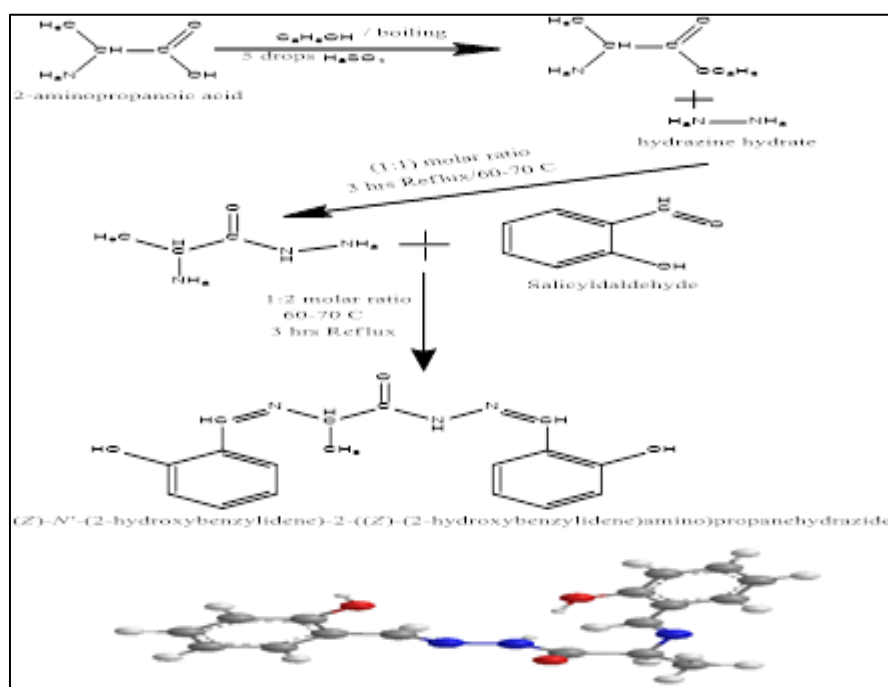


Figure 1: Chemical and 3D Structures of the prepared [**H₃L**] Ligand.

2.2 Metal complex preparation (2-20)

Metal complexes (**2–20**) were synthesized using a **1L:2M molar ratio** by refluxing the ligand with the corresponding metal salts in ethanol for **2 h at 80 °C** under continuous stirring. The resulting precipitates were filtered, washed, and dried over CaCl_2 to obtain the desired metal complexes. For the **BaSO₄ complex (11)**, five drops of HCl were added during the reaction. The synthesized complexes are presented in **Figure 2.2.3**.

2.3 Instruments and measurements

The synthesized compounds were characterized by **elemental analysis, TLC, FT-IR, ¹H-NMR, MS, UV-Vis, TGA/DTA, molar conductivity, magnetic susceptibility, and ESR spectroscopy**. FT-IR and ¹H-NMR analyses were performed using **JASCO FT/IR-300E** and **Bruker 350 MHz** instruments, respectively, while thermal behavior was evaluated using a **Shimadzu DT-30** analyzer. Magnetic properties and molar conductivity were measured by standard methods, and solid-state ESR spectra were recorded using a **Varian E-109 spectrometer with DPPH** as the reference.

2.4 Characterization via transmission electron microscopy (TEM)

Transmission electron microscopy (TEM) was employed to determine the **morphology, particle size, and distribution** of the ligand and its metal complexes. Colloidal suspensions were prepared in distilled water and deposited onto carbon-coated TEM grids, followed by air drying. The samples were then examined using a **JEOL 1230 TEM operated at 120 kV**.

2.5 Biological studies

2.5.1 In vitro study

The **in vitro cytotoxicity** of the synthesized compounds was assessed using the **Sulforhodamine B (SRB) assay**. Cancer cells were seeded in 96-well plates, treated with different concentrations of the compounds in triplicate, and incubated for **48 h at 37 °C under 5% CO₂**. Cells were then fixed, stained with SRB, and the bound dye was dissolved in Tris-EDTA buffer. Absorbance was measured using an **ELISA reader**, and the survival fraction was calculated to determine the cytotoxic activity of the tested compounds.

2.6 Study of toxicity

2.6.1 Animals

Forty male Wistar albino rats (170–200 g) were obtained from the **National Cancer Institute (NCI-Egypt)** and housed under controlled conditions at **23–26 °C**, with adequate ventilation and a **10 h light/14 h dark** photoperiod. The animals were provided with a standard laboratory diet containing **12% bran, 50% ground corn meal, 12% corn oil, 19% ground beans, 4% casein, 2% minerals, and 1% vitamins**, along with purified water ad libitum. The diet was supplied by **Meladco Feed, Cairo, Egypt**.

2.6.2 Acute toxicity

Sixteen animals were used to determine the LD₅₀ values of the tested compounds. The LD₅₀ was evaluated to establish the relationship between the administered dose and mortality, providing an indicator of the compounds' acute toxicity and potency. The determination was performed according to the method described by Akhila et al. [12].

2.6.3 Experimental design

Twenty-four male Wistar albino rats (185 ± 15 g) were allowed to acclimatize and then randomly divided into three groups (n = 8). **Group I** served as the healthy control, **Group II** received the Cu(II) complex at a dose of 1×10^{-5} mmol/L for 6 weeks, and **Group III** received the Zn(II) complex at the same dose for 6 weeks.

2.6.4 Blood collection samples

Following the experimental period, all animals were fasted overnight and euthanized under light ether anesthesia. Blood samples were collected from the inferior vena cava and centrifuged at **4,000 rpm for 5–10 min**. The obtained serum/plasma and **EDTA-anticoagulated whole blood** were collected for subsequent **hematological and biochemical analyses**.

2.6.5 Biochemical analyses:

Liver enzymes, Aspartate Aminotransferase (AST) and Alanine Aminotransferase (ALT) were measured via laboratory kits purchased from Human Diagnostic Kinetic Kits, Germany [13]. Serum albumin, blood urea, and serum creatinine levels were quantitatively determined using commercial diagnostic kits obtained from Diamond Diagnostics, Egypt, following the manufacturer's operational protocols [14].

2.6.6 Hematological analyses:

The complete blood count (CBC) containing Hb% hemoglobin was measured via Drabkin's solution (prepared), and white blood cells (WBCs) or total leucocyte (TLC) counts, red blood cells (RBCs), and platelets (PLTs) counts were measured manually [15].

2.7 Statistical analysis

Statistical analysis was performed using one-way ANOVA, followed by Duncan's multiple range test as a post hoc analysis. Data were analyzed using SPSS version 20.0 and expressed as mean $\pm$ SD. Differences were considered statistically significant at $P \leq 0.05$ [16].

3 RESULTS AND DISCUSSION:

3.1 Physical properties

For a long time (a year), the ligand and its metal complexes were crystalline, nonhydrophilic, and pigmented when exposed in air at room temperature (R.T 25 °C). These substances were insoluble in ethanol, water, benzene, methanol, and acetonitrile but soluble in dimethylsulfoxide (DMSO), dimethylformamide (DMF), nitromethane, nitrobenzene, and acetonitrile, chloroform and toluene. The chemical and physical analytical data shown in Table (1). Many effort have been made to prepare single or more crystals but unluckily. These efforts had failed. All acquired data fixed that, the formed (1 L: 2 M) molar ratio for all complexes.

Table 1: Ligand [HL] and its Metal Complexes, Physical Data and Analytical Data.

No	Ligand/Complexes	Color	FW	M.P (°C)	Yield (%)	Analysis /Found (Calculated.) (%)			
						C	H	N	M
(1)	[H ₃ L] C ₁₇ H ₁₇ N ₃ O ₃	Greenish blue	311	350	84	65.68 (64.90)	5.50 (5.30)	13.50 (13.45)	–
(2)	[(H ₃ L)(Cu) ₂ (CH ₃ COO) ₄ (H ₂ O) ₂].2H ₂ O C ₂₅ H ₃₃ N ₃ O ₁₄ Cu ₂	green	809	>300	81	41.1 (40.9)	5.8 (5.21)	5.78 (5.55)	17.49 (17.10)
(3)	[(H ₃ L)(Mn) ₂ (CH ₃ COO) ₄ (H ₂ O) ₂]. 2H ₂ O C ₂₅ H ₃₇ N ₃ O ₁₄ Mn ₂	Off white	739	>300	85	42.13 (41.8)	5.18 (4.8)	5.92 (5.10)	15.49 (14.10)
(4)	[(H ₃ L)(Ni) ₂ (CH ₃ COO) ₄ (H ₂ O) ₂]. 2H ₂ O C ₂₅ H ₃₇ N ₃ O ₁₄ Ni ₂	greenish	751	>300	84	41.18 (41.00)	5.89 (4.25)	9.46 (9.50)	14.37 (14.30)
(5)	[(H ₃ L)(Zn) ₂ (CH ₃ COO) ₄ (H ₂ O) ₂].2H ₂ O C ₂₅ H ₃₄ N ₃ O ₁₄ Zn ₂	Pale yellow	756	>300	78	41.12 (40.12)	4.55 (4.30)	9.75 (9.54)	15.9 (15.12)
(6)	[(H ₃ L)(Mg) ₂ (CH ₃ COO) ₄ (H ₂ O) ₂].2H ₂ O C ₂₅ H ₃₅ N ₃ O ₁₄ Mg ₂	Orange	709	>300	79	45.33 (45.23)	5.13 (4.52)	9.48 (9.85)	7.50 (6.14)
(7)	[(H ₃ L)(Co) ₂ (CH ₃ COO) ₄ (H ₂ O) ₂]. 2H ₂ O C ₂₅ H ₃₄ N ₃ O ₁₄ Co ₂	Reddish	744	>300	84	40.22 (40.12)	4.09 (4.15)	8.58 (8.65)	22.81 (23)
(8)	[(H ₃ L)(Cu) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 2H ₂ O C ₁₇ H ₂₆ N ₃ O ₁₆ S ₂ Cu ₂	Pale Blue Greenish	746	>300	79	28.41 (28.13)	3.51 (3.0)	8.85 (8.16)	13.69 (13.52)
(9)	[(H ₃ L)(Ni) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 2H ₂ O C ₁₇ H ₃₁ N ₃ O ₁₈ S ₂ Ni ₂	Dark Green	775	>300	82	27.3 (27.12)	4.15 (4.12)	5.6 (5.23)	15.8 (15.3)
(10)	[(H ₃ L)(Co) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 2H ₂ O C ₁₇ H ₂₅ N ₃ O ₁₆ S ₂ Co ₂	Orange	742	>300	84	25.78 (25.32)	3.55 (2.95)	5.68 (5.23)	11.62 (11.12)
(11)	[(H ₃ L)(Ba) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 2H ₂ O C ₁₇ H ₂₅ N ₃ O ₁₆ S ₂ Ba ₂	Dark yellowish	851	>300	81	23.72 (24.01)	3.13 (3.54)	4.84 (4.96)	31.64 (32.15)
(12)	[(H ₂ L)(Fe) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 4H ₂ O C ₁₇ H ₂₅ N ₃ O ₁₆ S ₂ Fe ₂	Green	767	>300	77	26.75 (26.23)	4.1 (3.95)	5.54 (5.4)	14.8 (14.3)
(13)	[(H ₃ L)(Mg) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 2H ₂ O C ₁₇ H ₂₅ N ₃ O ₁₆ S ₂ Mg ₂	Dark yellow	675	>300	82	29.57 (30.12)	2.91 (2.95)	8.85 (8.45)	31.71 (31.62)
(14)	[(H ₂ L)(Al) ₂ (SO ₄) ₂ (H ₂ O) ₄]. 2H ₂ O C ₁₇ H ₂₅ N ₃ O ₁₆ S ₂ Al ₂	Grayish	679	>300	75	31.53 (31.86)	4.20 (4.56)	6.49 (6.89)	8.33 (8.56)

(15)	$[(H_3L)(Zn)_2(SO_4)_2(H_2O)_4] \cdot 4H_2O$ $C_{17}H_{25}N_3O_{16}S_2Zn_2$	Pale green bluish	783	>300	81	28.19 (28.45)	3.76 (4.02)	5.80 (6.23)	18.05 (18.54)
(16)	$[(H_3L)(Ni)_2(NO_3)_4(H_2O)_3] \cdot 3H_2O$ $C_{17}H_{21}N_7O_{18}Ni_2$	Greenish	777	>300	85	27.51 (27.01)	2.8 (2.9)	13.2 (13.7)	20.61 (20.01)
(17)	$[(H_2L)(Fe)_2(NO_3)_4(H_2O)_3] \cdot 3H_2O$ $C_{17}H_{21}N_7O_{18}Fe_2$	Light bluish	769	>300	84	28.24 (28.12)	2.93 (2.85)	7.56 (7.95)	18.45 (19.01)
(18)	$[(H_3L)(Cd)_2(NO_3)_4(H_2O)_3] \cdot 3H_2O$ $C_{17}H_{27}N_7O_{18}Cd_2$	Offwhite	863	>300	76	23.8 (23.32)	3.1 (2.85)	11.4 (11.05)	26.11 (25.82)
(19)	$[(H_3L)(Mg)_2(CO_3)_2(H_2O)_4] \cdot 2H_2O$ $C_{19}H_{26}N_3O_{14}Mg_2$	Brownish	604	>300	81	40.03 (39.54)	4.77 (4.52)	7.37 (7.12)	8.53 (8.65)
(20)	$[(H_3L)(Cu)_2(CO_3)_2(H_2O)_4] \cdot 4H_2O$ $C_{19}H_{34}N_3O_{18}Cu_2$	Greenish	780	>300	84	28.5 (28.3)	4.25 (4.13)	5.25 (5.23)	15.88 (15.37)

3.2 IR spectra of the ligand [H₃L] (1) and its metal complexes

The FT-IR spectral analysis of the free ligand and its metal complexes (Table 2, Figures 2 and 3) elucidated the coordination mode and structural features. The spectrum of the free ligand displayed characteristic bands at 3450, 3373, 3190, 1700, and 1650 cm^{-1} , assigned to $\nu(OH)$, $\nu(N-H)$, $\nu(C=N)$, and $\nu(C=O)$ stretching vibrations, respectively, along with broad bands (3600–2670 cm^{-1}) indicating intra- and intermolecular hydrogen bonding. Upon complexation, the shift of $\nu(C=N)$ and $\nu(C=O)$ to lower frequency regions (1675–1570 cm^{-1} and 1682–1610 cm^{-1}) confirmed the coordination via azomethine nitrogen and carbonyl oxygen atoms, further evidenced by the emergence of new non-ligand bands at 610–640 cm^{-1} ($\nu(M-N)$) and 520–580 cm^{-1} ($\nu(M-O)$). Additionally, broad absorption in the 3570–3100 cm^{-1} region confirmed the presence of hydrated or coordinated water, while characteristic bands at ~1220, 800, and 360 cm^{-1} was attributed to inorganic anions (NO_3^-/CO_3^{2-}), alongside the $\nu(S=O)$ vibration of coordinated DMSO at ~1030 cm^{-1} [17].

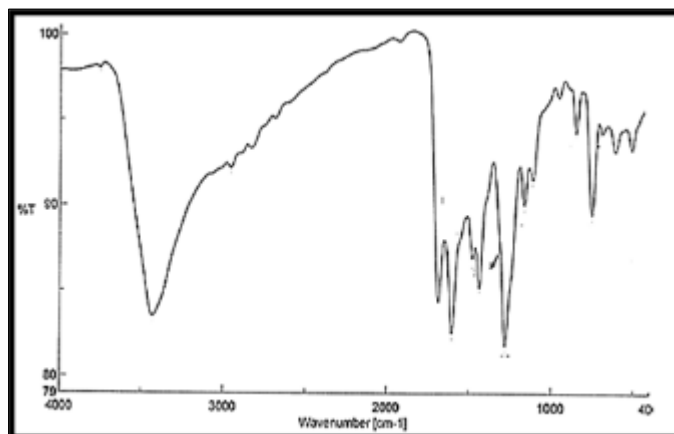


Figure 2: IR spectrum of the Ligand

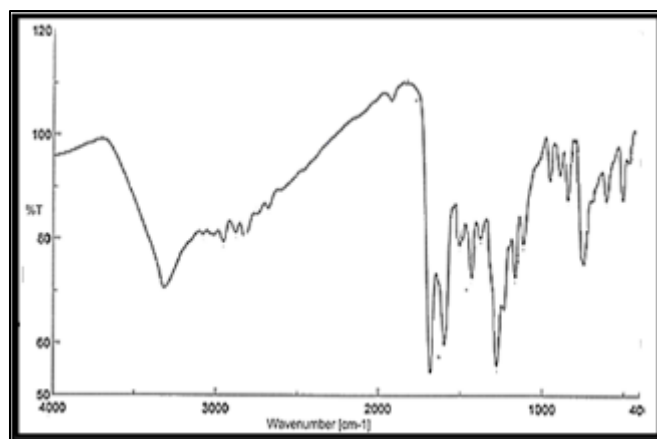


Figure 3: IR spectrum of the Cu(II) complex

Table 2: IR Frequencies Bands (cm⁻¹) of the Ligand and its Metal Complexes.

No.	$\nu(\text{H}_2\text{O})$	$\nu(\text{O H})$	$\nu(\text{H-bondings})$	$\nu(\text{NH})$	$\nu(\text{C=O})$	$\nu(\text{C=N})$	$\nu(\text{Ar})$	$\nu(\text{OAc})/\text{SO}_4$	$\nu(\text{M-O})$	$\nu(\text{M-N})$
(1)	3370-3220	3450, 3373	3600-3320, 3380-2670	3190	1700	1650	1540, 1450, 818, 750	-	-	-
(2)	3540-3250	3413, 3365	3620-3320, 3210-2774	3175	1675	1610	1510, 1435, 850, 770	1240, 1160, 1110, 750	610	580
(3)	3570-3280, 3350-3225	3420, 3372	3610-3320, 3319-2830	3160	1680	1615	1510, 1435, 850, 770	1285, 1180, 1080, 750	640	577
(4)	3560-3385, 3310-3185	3410, 3320	3610-3190, 3180-2680	3180	1610	1616	1470, 1438, 852, 753	1252, 1165, 1082, 755	635	583
(5)	3560-3370, 3320-3180	3420, 3379	3600-3210, 3200-2700	3150	1610	1635	1486, 1430, 850, 765	1260, 1125, 1190, 735	610	516
(6)	3340-3160	3415, 3380	3560-3210, 3200-2720	3170	1651	1610	1540, 1515, 850, 768	1230, 1075, 1110, 770	615	513
(7)	3350-3185	3400, 3385	3645-3260, 3210-3260	3187	1680	1605	1470, 1438, 852, 753	1260, 1260, 1093, 690	640	520
(8)	3370-3130	3420, 3375	3650-3280, 3270-2650	3155	1615	1610	1486, 1430, 850, 765	1430, 1340	615	525
(9)	3510-3350, 3315, 3170	3418, 3245	3610-3330, 3310-2870	3180	1614	1612	1470, 1438, 852, 753	1354, 1461	610	530
(10)	3325, 3100	3422	3650-3330, 3320-2675	3178	1610	1615	1540, 1515, 850, 768	1465, 1340	630	537

(11)	3300,3150	3415	3640-3325, 3320-2870	3160	1682	1650	1486,143 0,850,76 5	1480,1360	610	520
(12)	3320-3165	3420	3650-3350, 3340-2670	3170	1655	1670	1540,151 5, 850,768	1470,1355	618	560
(13)	3325-3160	3415	3550-3340, 3340-2650	3150	1681	1664	1540,151 0, 850,768	1430,1360	618	580
(14)	3365-3260	3432 ,338 0	3610-3340, 3340-2675	3170	1676	1660	1540,151 0, 850,768	1420,1210	615	515
(15)	3325-3160	3418	3650-3340, 3340-2610	3160	1670	1676	1540,151 0, 850,768	1430,1240	610	510
(16)	3325-3180	3425	3620-3340, 3340-2610	3180	1650	1665	1540,151 0, 850,768	-	618	520
(17)	3300-3170	3320 ,330 0	3600-3320 3300-2700	3142	1670	1580	1550, 760	-	610	510
(18)	3340-3180	3410 3100	3540-3320 3310-2800	3150	1640	1570	1560, 754	-	615	580
(19)	3300-3170	3450 3110	3570-3340, 3300-2900	3150	1670	1610	1560, 850	-	618	570
(20)	3320-3180	3400 3100	3550-3300	3155	1675	1610	1555,765	-	618	520

3.3 ^1H -NMR spectra

The ^1H -NMR spectra of the ligand and its diamagnetic Zn(II) and Mg(II) complexes in DMSO- d_6 confirmed the proposed structures. The free ligand showed characteristic signals for $-\text{CH}_3$, $-\text{CH}=\text{}$, and hydrazide $-\text{NH}-\text{N}=\text{C}-$ protons at 2.0/3.7, 5.4, and 8.9 ppm, respectively, with aromatic protons at 6.5–8.3 ppm. Upon complexation, the $-\text{NH}-$ and $-\text{CH}=\text{}$ signals shifted upfield to 8.0–8.3 and 4.8–5.1 ppm, respectively, confirming coordination of the **azomethine nitrogen to the metal center (M $\leftarrow$ N)**.

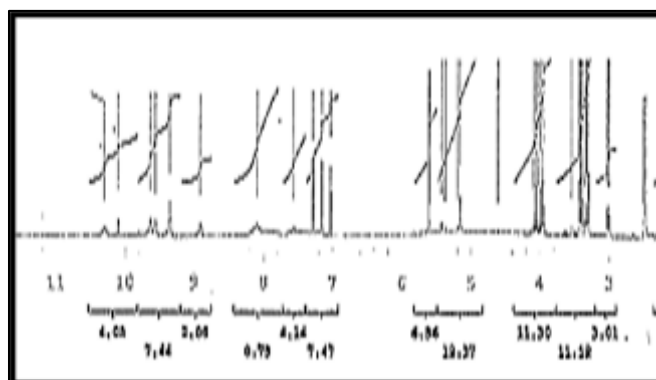


Figure 4: ^1H -NMR of the ligand

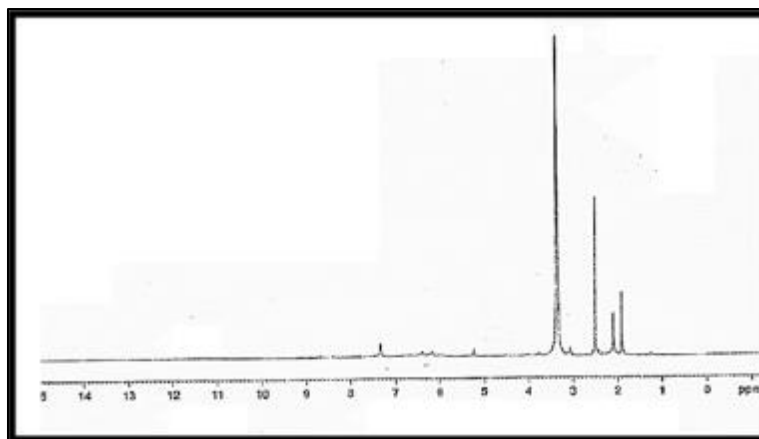


Figure 5: ^1H -NMR of the Zn(II) complex

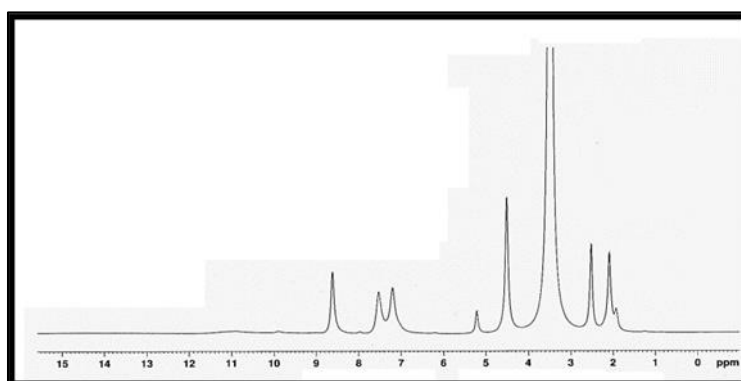


Figure 6: ^1H -NMR of the Mg^{II} complex

3.4 Conductance measurements

Molar conductance in various solvents of seven metal complexes (1×10^{-3}) was carried out to show the effect of solvent. The complexes displayed low molar conductivities as shown in table (3). Together the ligand and its metal complexes had non-electrolytic properties [18]. As shown as in figures 7 - 10 respectively.

Table 3: Molar conductance in various solvents

Complex No	DMSO	DMF	Acetonitrile	Nitrobenzene	Acetone	Nitromethan
Complex (2)	18	9.3	50	11	10	23
Complex (3)	20	10.7	57	14	12	24
Complex (4)	17	11.5	53	12	9.8	22
Complex (5)	15.8	8.7	48	11.7	9.5	20
Complex (6)	16	8	46	10.2	8.5	19
Complex (7)	16.8	10.2	54	12	10	22
Complex (8)	21	17	62	16	7.8	27

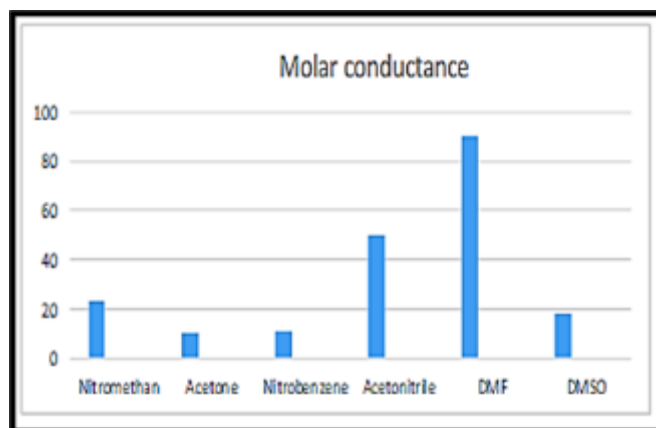


Figure 7: Molar conductance for complex (2)

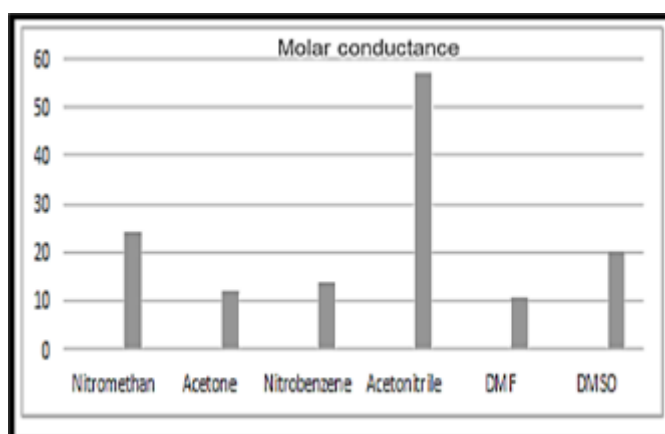


Figure 8: Molar conductance for complex (3)

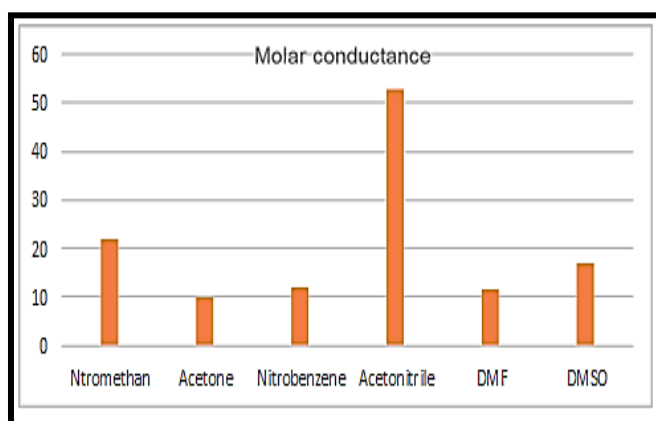


Figure 9: Molar conductance for complex (4)

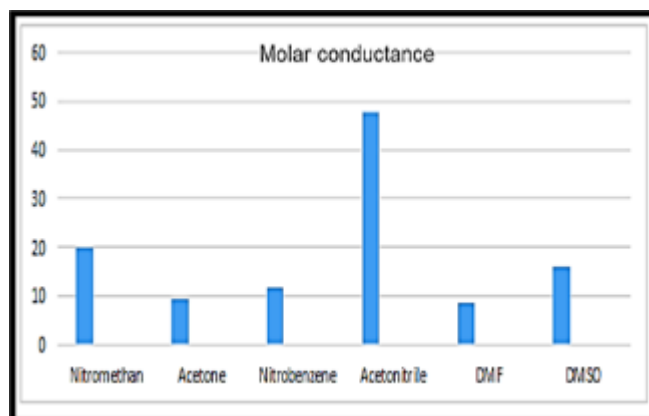


Figure 10: Molar conductance for complex (5)

3.5 Mass spectra

The mass spectrum of the ligand showed a molecular ion peak at $m/z = 322$, consistent with its calculated molecular mass and supporting the proposed structure. The characteristic fragment peaks at $m/z = 54, 63, 90, 119, 221, 238$, and 309 further confirmed the ligand framework. Similarly, the Zn(II) complex (5) exhibited a molecular ion peak at $m/z = 728$, in agreement with its calculated molecular mass. The characteristic fragmentation peaks at $m/z = 54.89, 219, 238, 509$, and 548 provided additional support for the proposed structure. The mass spectra of the ligand and Zn(II) complex (5) are shown in Figures 11 and 12, respectively.

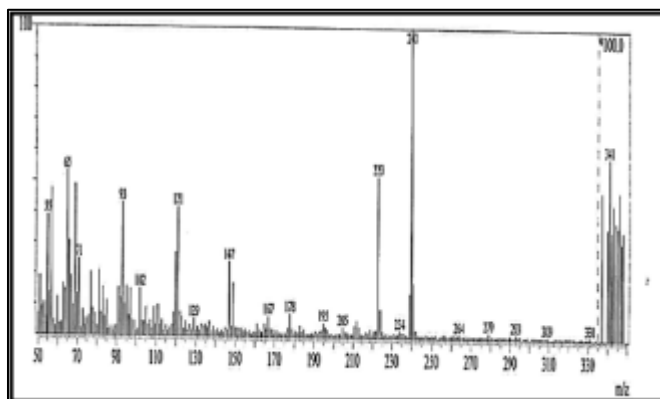


Figure 11: Mass spectrum of the ligand

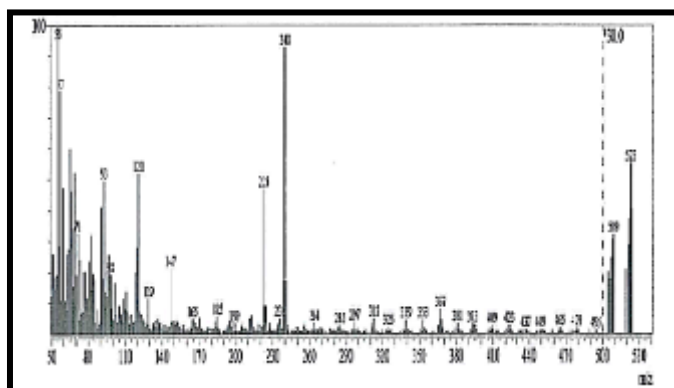


Figure 12: Mass spectrum of the Zn(II)

3.6 Electronic spectra

The electronic absorption spectrum of free [HL] exhibited three characteristic bands at 280, 310, and 350 nm ($\pi \rightarrow \pi^*$, $n \rightarrow \pi^*$, and CT transitions), while its metal complexes displayed corresponding intraligand transitions alongside diagnostic d–d bands [20]. Specifically, Cu II complexes (**2**, **8**, **20**) exhibited d–d absorption in the 430–620 nm region with magnetic moments of 1.69–1.71 B.M., indicating a Jahn-Teller distorted octahedral geometry (d^9). NiII complexes (**4**, **9**, **16**) displayed d–d transitions at 440–745 nm with magnetic moments of 3.01–3.14 B.M., supporting a distorted octahedral environment ($\nu_2/\nu_1=1.59$ –1.64). Furthermore, MnII, CoII ($\mu_{\text{eff}}=4.48$ –4.80 B.M.), and FeIII ($\mu_{\text{eff}}=5.80$ –6.12 B.M.) complexes revealed characteristic high-spin octahedral geometries, whereas the remaining diamagnetic complexes were dominated exclusively by intraligand transitions [21].

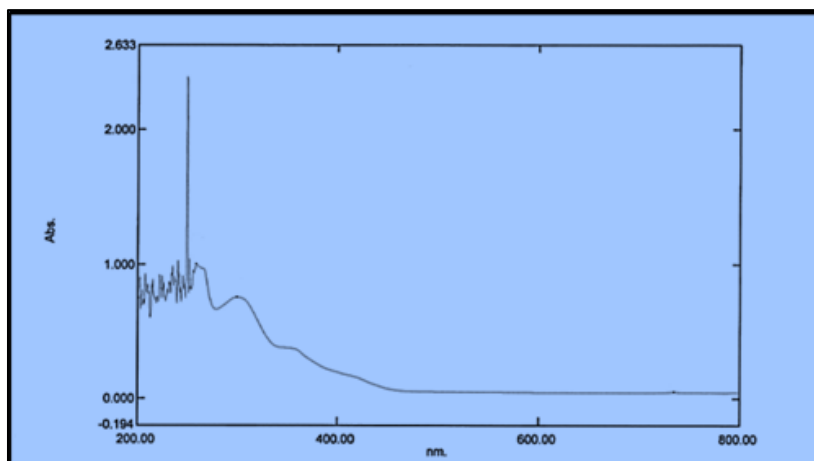


Figure 13: Electronic spectrum of the Zn II

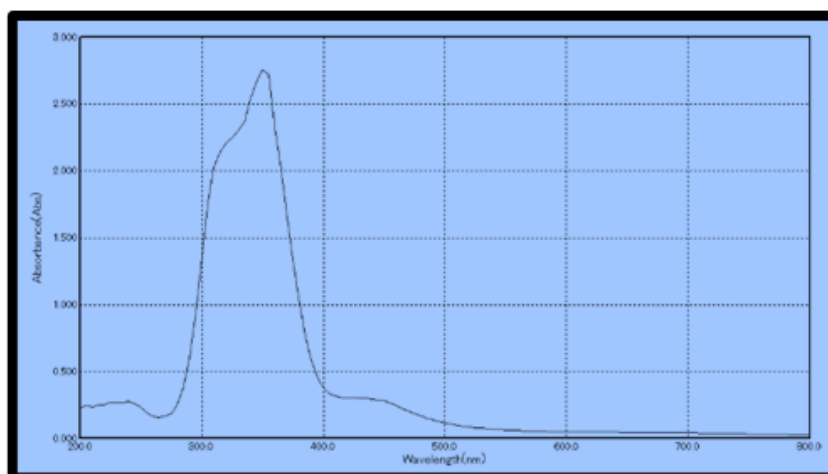


Figure 14: Electronic spectrum of the Cu II

Table 4: B.M., the prepared compounds' magnetic moments, and electronic absorption spectral bands (nm).

Comp. No.	λ_{max} (nm)	μ_{eff}
(1)	280, 310, 350	-
(2)	250, 310, 320, 450, 550, 620	1.71
(3)	260, 310, 410, 490, 580, 615	5.65
(4)	260, 310, 440, 560, 620, 720	3.01
(5)	250, 310, 300	Diamagnetic
(6)	250, 310, 320	Diamagnetic
(7)	260, 310, 380, 410, 580, 610	4.48

(8)	260, 310, 450, 520, 610	1.69
(9)	275, 320, 410, 450, 530, 610, 740	3.16
(10)	270, 310, 440, 570, 625	4.80
(11)	260, 320, 310	Diamagnetic
(12)	260, 310, 460, 520, 620	5.80
(13)	250, 270, 310	Diamagnetic
(14)	260, 280, 310	Diamagnetic
(15)	260, 300, 310	Diamagnetic
(16)	260, 310, 460, 540, 600, 730	3.4
(17)	260, 310, 450, 560, 620	6.12
(18)	270, 310, 300	Diamagnetic
(19)	270, 310, 300	Diamagnetic
(20)	260, 300, 310, 430, 560, 615	1.71

3.7 Thermal analysis [22]

Thermal stability and decomposition pathways of representative complexes (2, 3, 12, 18, and 20) were investigated by TG/DTA under a nitrogen atmosphere (Table 5). The thermograms showed a multistep decomposition pattern. The initial endothermic event at 40–50°C was attributed to hydrogen-bond cleavage, followed by the loss of lattice water below 85°C. Further endothermic steps at 125–140°C and 160–250°C corresponded to the loss of coordinated water and coordinated anions, respectively. The experimental weight losses were in good agreement with the calculated values. Subsequent thermal decomposition at higher temperatures involved degradation of the organic ligand framework, ultimately leaving the corresponding metal oxides (CuO, MnO, NiO, FeO, or CdO) as stable residues. These results confirm the good thermal stability and proposed compositions of the investigated complexes.

Table 5: Thermal analyses of some Metal complexes

Compound No.	Temp. (°C)	DTA (peak %)		TGA (Wt.loss %)		Assignments
		Endo	Exo	Calculated.	Found	
(2)	45	Yes	No	0	0	Broken of H-bondingss
	70	Yes	No	2.46	2.30	Loss of (H ₂ O) hydrated water
	125	Yes	No	7.58	7.45	Loss of (3H ₂ O) molecules
	160	Yes	No	25.40	23.9	Loss of (2OAC) group
	320	No	Yes	0	0	Melting point
(3)	570 460,510,540,	No	Yes	37.7	37.4	Decomposition process with the formation of 2CuO
	50	Yes	No	0	0	Broken of H-bondingss
	85	Yes	No	2.52	2.41	Loss of (H ₂ O) hydrated water molecule
	135	Yes	No	7.8	7.65	Loss of (3H ₂ O) Coordinated water
	185	Yes	No	36.76	36.52	Loss of 4(OAC) group
	360	No	Yes	0	0	Melting point
	400,440,475,560,570	No	Yes	35.1	34.82	Decomposition process with the formation of 2Mn
	40	Yes	No	0	0	Broken of H-bondingss

(9)	80	Yes	No	8.2	8.1	Loss of (2H ₂ O) hydrated water molecules
	140	Yes	No	12.69	12.13	Loss of (5H ₂ O) Coordinated water molecules
	250	Yes	No	31.02	30.92	Loss of 2SO ₄ group
	340	No	Yes	0	0	Melting point
	420,440,480, 550,580	No	Yes	35.13	35.0	Decomposition process with the formation of 2NiO
(12)	45	Yes	No	0	0	Broken of H-bondingss
	70	Yes	No	4.75	4.56	Loss of (2H ₂ O) hydrated water molecules
	130	Yes	No	12.45	12.21	Loss of (5H ₂ O) coordinated water molecule
	240	Yes	No	30.38	30.12	Loss of 2SO ₄ group
	340	No	Yes	0	0	Melting point
	410,510,540,600	No	Yes	32.72	32.51	Decomposition process with the formation of 2FeO
(18)	45	Yes	No	0	0	Broken of H-bondingss
	75	Yes	No	4.2	3.95	Loss of(2H ₂ O) hydrated water molecules
	140	Yes	No	6.6	6.52	Loss of(3 H ₂ O)Coordinate water molecules
	235	Yes	No	32.3	32.0	Loss of(4NO ₃)
	320	No	Yes	0	0	Melting point
	340,370,440,510,580	No	Yes	18.1	17.98	Decomposition process with the formation of 2CdO
(20)	40	Yes	No	0	0	Broken of H-bondingss
	70	Yes	No	5.75	5.54	Loss of (3H ₂ O) hydrated water molecules
	140	Yes	No	12.1	11.98	Loss of (5H ₂ O) coordinated water molecules
	210	Yes		17.9	`17.7	Loss of (2CO ₃) group
	370	No	Yes	0	0	Melting point
	380,440,510,540,610	No	Yes	23.69	23.35	Decomposition process with the formation of 2CuO

3.8 Electron spin resonance (ESR):

The solid-state ESR spectra provided key insights into the coordination geometries and bonding characteristics of the metal complexes. The CuII complexes (2, 8, 20) exhibited axial spectra with $g_{\parallel} > g_{\perp} > 2.0024$, confirming a $d_{x^2-y^2}$ (2B_{1g}) ground state in a tetragonally distorted octahedral environment ($g_{\parallel}/A_{\parallel} = 153.4\text{--}179.0$ cm). The exchange parameter ($G \geq 4.0$) indicated negligible spin-exchange coupling between copper centers. Evaluated bonding parameters ($K < 1.0$, $g_{\parallel} < 2.31$, $\alpha^2 = 0.60\text{--}0.71$, and $\beta_1^2 = 0.88\text{--}0.94$) demonstrated significant in-plane σ - and π -covalency, whereas out-of-plane π -interaction ($\beta^2 > 1.0$) was predominantly ionic, aligned with calculated %ad² values (52.5–64.9%). In contrast, the MnII, CoII, and FeIII complexes displayed isotropic signals ($g_{\text{iso}} = 2.001\text{--}2.050$), characteristic of distorted octahedral geometries [23].

Table 6: Electron spin resonance data of some metal complexes

Complex	$g_{\parallel}$	$g_{\perp}$	g_{iso}^a	$A_{\parallel}$ (G)	$A_{\perp}$ (G)	A_{iso} b_o (G)	G^c	ΔE_x y (cm ⁻¹)	ΔE_x z (cm ⁻¹)	$K_{\perp}$ z	$K_{\parallel}$ z	K^2	K	$g_{\parallel}/A_{\parallel}$ (cm ⁻¹)	α 2	β^2	1β 2	-2β	a^2 d (%)
(2)	2.241	2.062	2.12	140	5.0	50	4.0	17543	22831	0.79	0.63	0.74	0.87	153.4	0.71	1.1	0.89	152.5	64.9
(3)	-	-	2.007	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
(8)	2.20	2.051	2.10	120	7.5	45	4.0	17699	22472	0.65	0.53	0.61	0.78	179	0.6	1.08	0.88	123	52.5
(11)	-	-	2.0011	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
(12)	-	-	2.003	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
(17)	-	-	2.05	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-
(20)	2.231	2.0551	2.112	125	5.0	45	4.18	17857	22989	0.73	0.61	0.69	0.83	171.5	0.65	1.12	0.94	133	56.7

Based on all the aforementioned studies, the proposed chemical structures of the investigated metal complexes are illustrated as follows

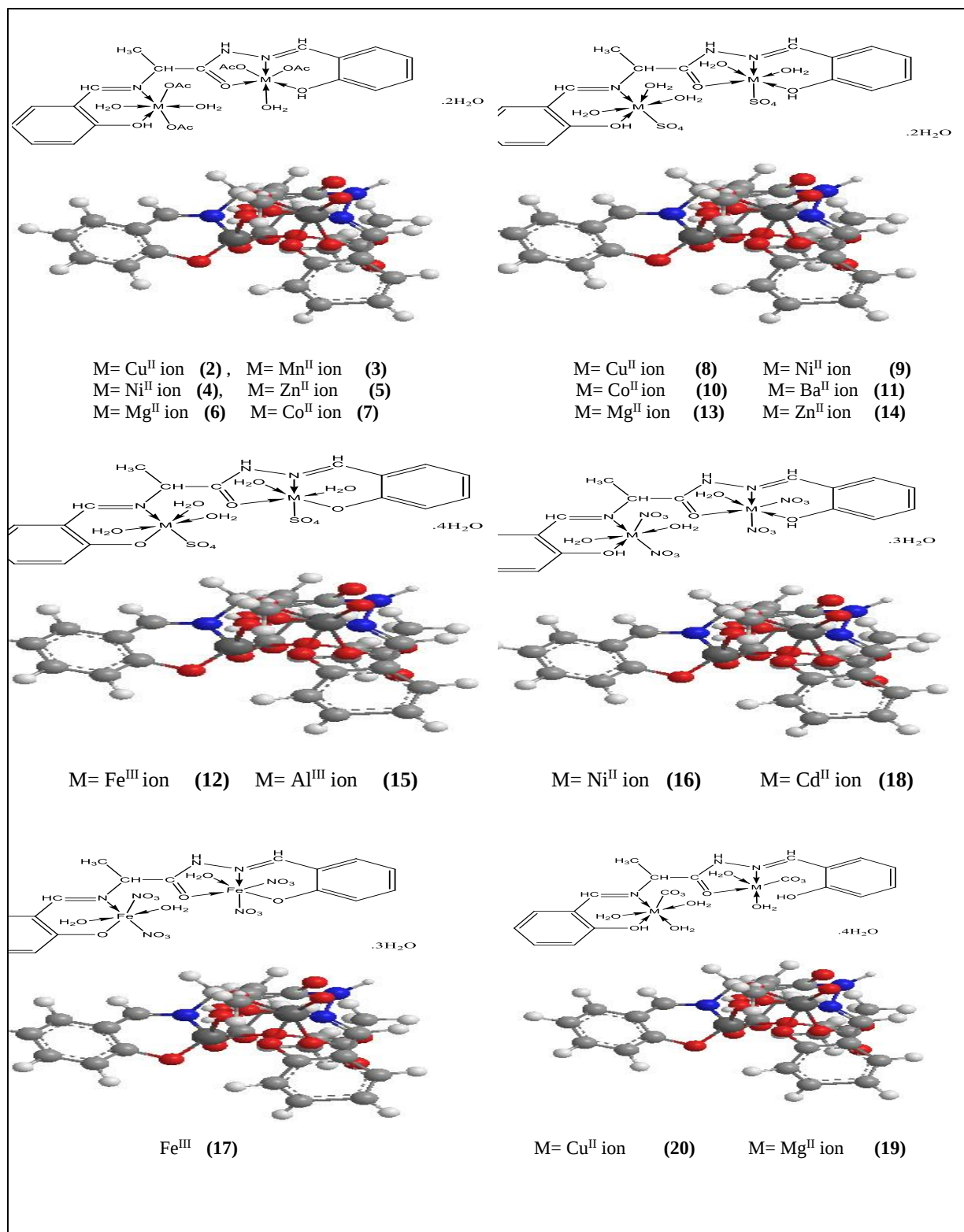


Figure 15: Chemical and 3D structures of the prepared metal complexes.

3.9 Transmission electron microscopy (TEM) characterization

Copper II complex(2) and Zinc II complex(5) complexes had average diameters of 21.875 ± 6.989 nm and 12.11 ± 1.336 nm, respectively. Each particle had a diameter of 1 to 100 nm and a size of a nanometer. These compounds were present as nanoparticles, as seen in Figures (3) and (4).

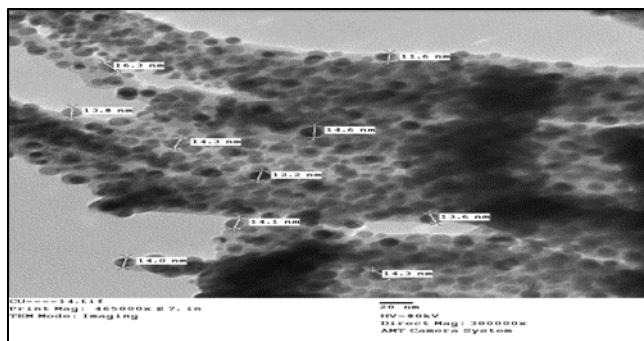


Figure 16: Copper II complex nanoparticles

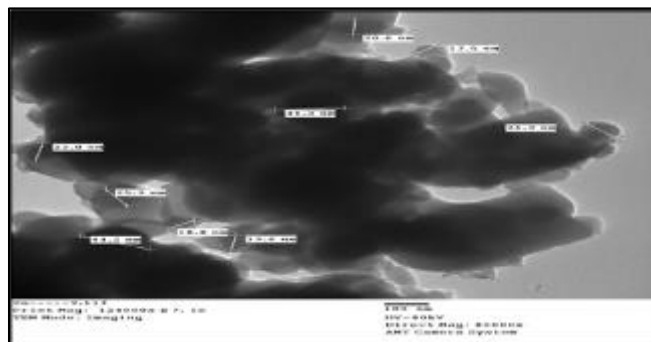


Figure 17: Zinc II complex nanoparticles

When compared to large-sized particles, metal complexes exhibited new or reinforcing size-dependent features for the material or compounds. The complex particles were created as nanosized particles, meaning their diameter ranged from 1 to 100 nm. They also offer numerous advantages and benefits, including a smaller dosage form, stable dosage forms, improved bioavailability, reduced toxicity, dose proportionality, and many more for a variety of medications. [24, 25].

3.10 In vitro cytotoxicity:

The cytotoxic activity of the ligand and its metal complexes (2), (5), and (7) was evaluated against human breast cancer (MCF-7) cells. All complexes showed greater anticancer activity than the free ligand, consistent with Tweedy's chelation theory. Among the tested compounds, Zn(II) complex (7) exhibited the highest potency, with an IC_{50} of 5.01 μ M, followed by complexes (2) and (5), with IC_{50} values of 30.0 and 31.6 μ M, respectively. The enhanced activity may be attributed to metal chelation, which modifies the electronic properties, lipophilicity, and DNA-binding ability of the complexes. The observed cytotoxicity may also involve ROS generation and oxidative stress, contributing to the dose-dependent reduction in MCF-7 cell viability [26].

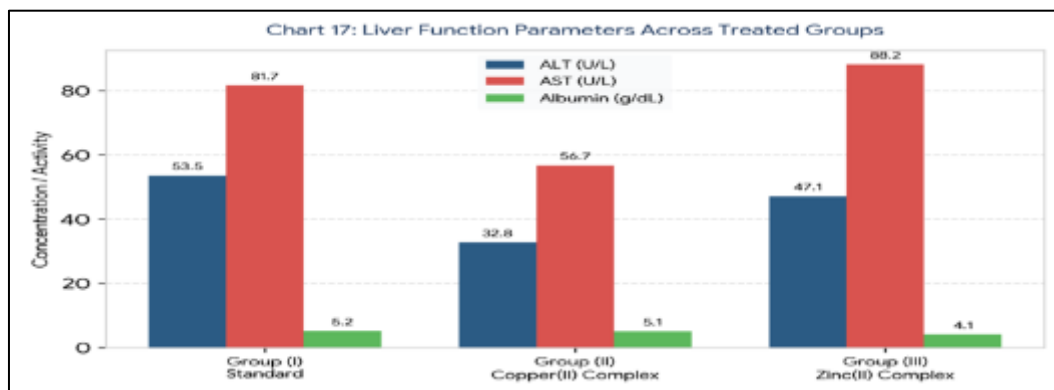
3.10.1 Overview of In Vivo Toxicity Assessments

As detailed in Table 10, liver function evaluation based on plasma albumin levels and key hepatic enzymes (ALT and AST) showed no statistically significant toxicological differences between the standard control group and the treated groups in normal rats. Furthermore, renal function markers (serum creatinine and blood urea nitrogen) as well as comprehensive hematological parameters (Table 11 and Table 12) exhibited no significant adverse variations following administration of Copper(II) and Zinc(II) complexes, confirming the biological safety profile of these coordination compounds at a dosage of 1×10^{-3} mmol/L.

Table 10: Statistical analysis (ANOVA) of liver function parameters among the experimental groups.

Enzymes / Parameters	Units	Group (I) Standard Control	Group (II) Copper(II) Complex	Group (III) Zinc(II) Complex
ALT (Alanine Aminotransferase)	U/L	53.538 ± 4.508	32.789 ± 5.190	47.142 ± 3.874
AST (Aspartate Aminotransferase)	U/L	81.651 ± 7.911	56.726 ± 4.240	88.211 ± 5.810
Albumin	g/dL	5.150 ± 2.197	5.113 ± 2.760	4.091 ± 1.229

Note: Values are expressed as Mean ± Standard Deviation (SD). Statistical analysis was performed using one-way ANOVA ($p < 0.05$). Means sharing the same superscript letters within a row are not significantly different from each other. Treatments were administered at a concentration of 1×10^{-5} mmol/L.

**Figure 18: Liver enzyme activities (ALT, AST) and albumin concentrations across experimental groups.****Table 11: Statistical analysis (ANOVA) of renal function tests among the experimental groups.**

Kidney Parameters	Units	Group (I) Standard Control	Group (II) Copper(II) Complex	Group (III) Zinc(II) Complex
Blood Urea	mg/dL	34.166 ± 3.594	31.819 ± 2.007	38.005 ± 3.101
Serum Creatinine	mg/dL	1.504 ± 0.199	1.499 ± 0.101	1.536 ± 0.100

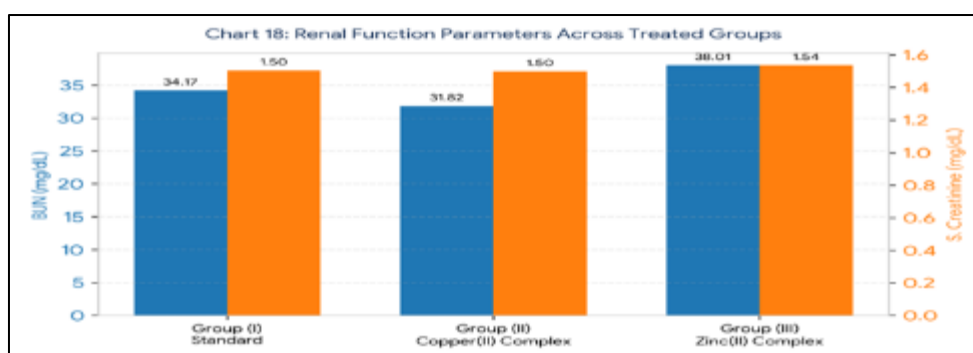
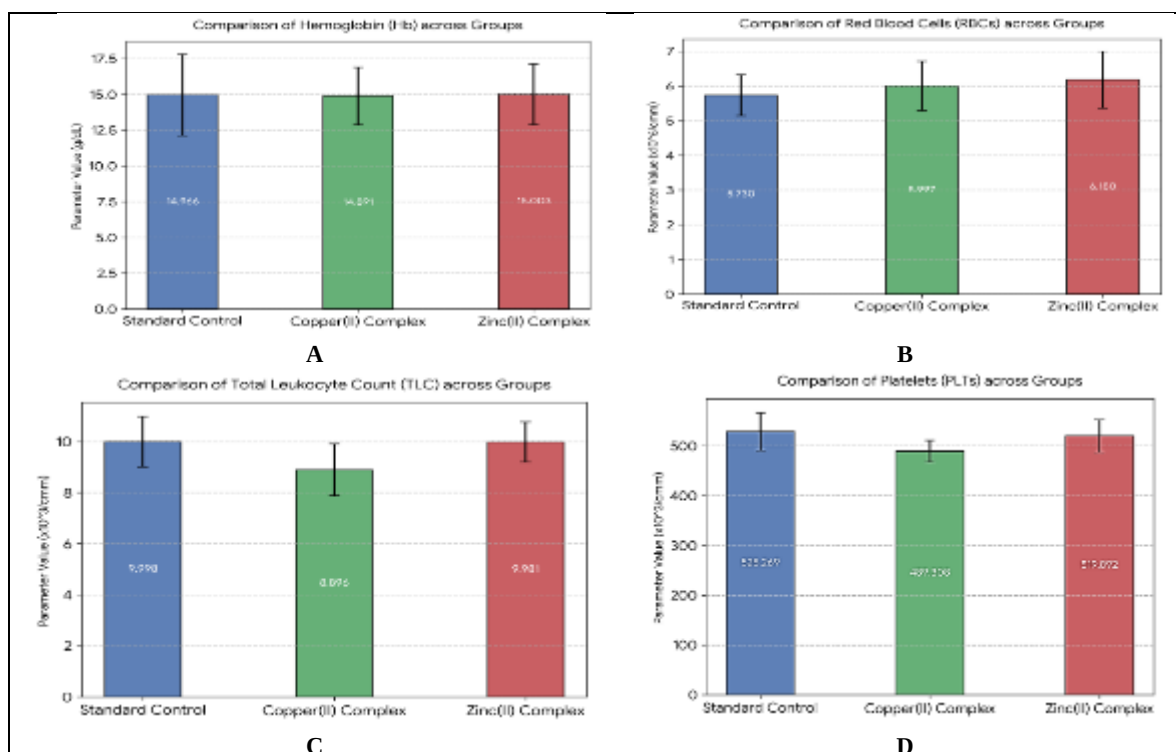
**Figure 19: Renal function parameters (B.Urea and Serum Creatinine) across experimental groups.**

Table 12: Statistical evaluation (ANOVA) of hematological parameters across experimental groups.

Hematological Parameter (CBC)	Units	Group (I) Standard Control	Group (II) Copper(II) Complex	Group (III) Zinc(II) Complex
Hemoglobin (Hb)	g/dL	14.966 ± 2.842	14.891 ± 2.012	15.003 ± 2.112
Red Blood Cells (RBCs)	×10 ⁶ /cmm	5.730 ± 0.591	5.997 ± 0.709	6.180 ± 0.819
Total Leukocyte Count (TLC)	×10 ³ /cmm	9.998 ± 0.983	8.896 ± 1.019	9.981 ± 0.783
Platelets (PLTs)	×10 ³ /cmm	528.269 ± 37.974	489.308 ± 21.891	519.892 ± 32.612

**Figure 20: A, B, C and D: Hematological parameters across experimental groups**

4 CONCLUSION

Nano-organometallic complexes of Fe(III), Mn(II), Co(II), Ni(II), Cu(II), Cd(II), Mg(II), Al(III), Zn(II), and Ba(II) with an amido-salicylaldehyde ligand were successfully synthesized and characterized using spectroscopic and thermal techniques. The complexes were predominantly octahedral and non-electrolytic, with considerable covalent character. Biological studies showed enhanced, dose-dependent cytotoxicity against MCF-7 breast cancer cells compared with the free ligand, in agreement with Tweedy's chelation theory [25]. Molecular docking further demonstrated favorable interactions with the target protein active sites, supporting the potential of these nano-organometallic complexes as promising anticancer candidates.

STATEMENTS ON COMPLIANCE WITH ETHICAL STANDARDS

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Disclosure of conflict of interest

The authors have declared no conflict of interest.

Statement of ethical approval

This study was performed after getting permission from the Institutional Animal Ethical Committee, Menoufia University, Egypt (approval ID: MUFS/S/CH/1/25).

Statement of informed consent

This study did not involve human participants, and therefore, informed consent was not required.

Author's Contributions

A. S. El-Tabl and M.M. Abd Elwahed designed the study and performed the complexes synthesis. A. M. Ashour, R. M. Mustafa and M. F. Mohamed performed most of the experiments. A. M. Ashour and S.M. Faheem analyzed and interpreted the data. A. S. El-Tabl and A.M. Ashour wrote the first version of the manuscript. All authors reviewed and approved the final version of the manuscript.

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