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Plant-based Natural inhibitors of human liver carcinogenesis: A mechanistic overview, focusing on hepatitis B and hepatitis C viruses

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Abstract

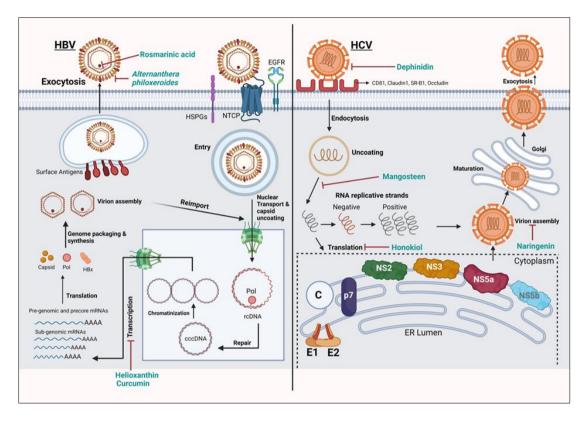
Hepatitis B and C viruses can lead to serious complications such as hepatic fibrosis, liver cirrhosis, and hepatocellular carcinoma (HCC) and are therefore responsible for a significant portion of liver cancer cases worldwide, with over 1.3 million deaths annually. The mechanisms by which hepatitis viruses contribute to HCC include DNA integration into the host genome, metabolic reprogramming, induction of the cellular stress response pathway, and interference with tumour suppressors. HBV is a DNA virus from the *Hepadnaviridae* family, and HCV is an RNA virus from the *Flaviviridae* family. Both viruses are transmitted through contact with infected bodily fluids, such as blood or sexual fluids. It is important to get tested for hepatitis B and C and to seek treatment as early as possible to prevent the progression to liver cancer. While there is a vaccine available for Hepatitis B, there is currently no vaccine for Hepatitis C. But some natural medicines have demonstrated antiviral activity against the hepatitis B and C viruses. Therefore, it is important to explore natural alternatives for the treatment of this disease. This review aims to summarise the pathogenesis of hepatitis B and C and their link to hepatocellular carcinoma, as well as to highlight natural compounds with the potential to treat hepatitis through various mechanisms at different stages of infection. These natural compounds may offer an alternative to chemical-based medications in the treatment and control of hepatitis by inhibiting or disrupting the entry, activity, or replication of the virus within the host.

Keywords: Hepatitis B; Hepatitis C; Human Liver Cancer; Hepatitis B virus associated Hepatocellular carcinoma; Natural inhibitors

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Graphical Abstract



Plant-based compounds and their mechanism of action against hepatitis B and C viruses.

1. Introduction

Infections with HBV and HCV are responsible for a significant portion of the liver disorders that are seen around the world. As a result of the fact that the two hepatotropic viruses share the same mode of transmission, co-infection with the two viruses is extremely common. This is especially true in areas of the world where HBV infection is more common, as well as in populations that are at a high risk for parenteral infection [1].

1.1. Hepatitis B Virus

Hepatitis B virus (HBV) is a member of the *Hepadnaviridae* family of viruses that infects exclusively the hepatocytes (liver cells) of humans and some non-human primates. It is found in several different forms in the blood, with the infectious form being known as the Dane particle. The Dane particle is a small, enveloped virus with a partially double-stranded DNA genome of approximately 3.2 kilobases in size and a diameter of 42nm, that is linked to a polymerase and surrounded by a nucleocapsid. It also contains three envelope proteins called the large (L), middle (M), and small (S) surface proteins, which contain domains essential for attachment to hepatocytes (Fig.1) [2]. The C-terminal S domain is common to all three envelope proteins, while the M protein also contains an extra N-terminal preS2 domain and the L protein contains a preS1 domain in addition to the preS2 and S domains [3]. In addition to the Dane particle, there are two other forms of HBV that are secreted in large amounts and known as subviral particles (SVPs) [4]. These SVPs contain only envelope proteins and are non-infectious. The SVPs can be either spherical or filamentous in shape, with the spherical SVPs being composed of S (90%) and M proteins (10%), and the filamentous SVPs containing S (80%), M, and L proteins (10% each) [5]. The lipid composition of SVPs has been determined to consist mainly of phosphatidylcholine, cholesteryl ester, cholesterol, and triglycerides, but the lipid composition of Dane particles has not yet been determined [6]. The relevance of SVPs in the life cycle of HBV is unclear, but it has been suggested that they may act as a decoy for the immune system, protecting the Dane particle from the neutralizing humoral response [7].

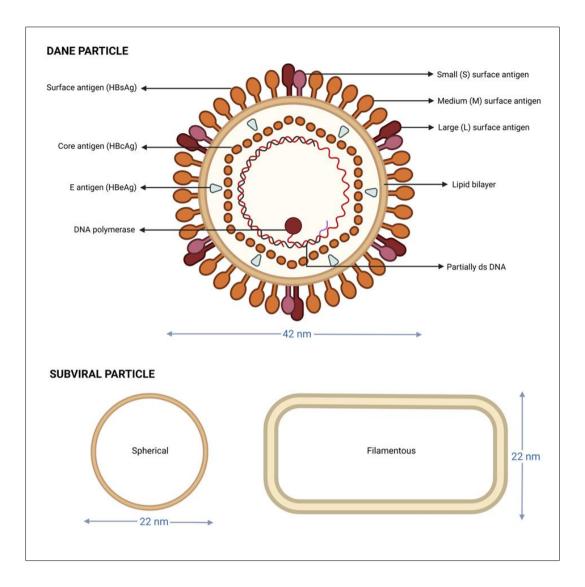


Figure 1 Schematic representation of Hepatitis B

1.1.1. Virus interaction with Hepatocytes

The hepatitis B virus (HBV) infects the liver and causes chronic liver disease by binding to liver cells via a protein called sodium taurocholate co-transporting polypeptide (NTCP) and then entering the cell to replicate **(Fig.2)**. The process of HBV entering a cell may involve the interaction with multiple receptors and may be a complex, multi-step process that involves endocytosis, a process by which the cell takes in molecules from outside the cell. It is thought that additional host factors, or proteins produced by the host cell, are required for susceptibility to HBV infection[8]. Some host factors that may play a role in HBV infection include the epidermal growth factor receptor (EGFR) and the protein E-cadherin [9]. The mechanism by which HBV gains access to the cell once it has interacted with its receptor and coreceptor is not fully understood, but some studies have suggested that HBV enters cells through a process called Caveolin-1-mediated endocytosis, while others have found evidence for the involvement of Clathrin-mediated endocytosis (CME) [10]. Inhibition of CME has been shown to decrease HBV infection in some studies [11, 12, 13]. Further understanding of the mechanisms involved in HBV entry into cells could lead to the development of new inhibitors to eliminate the virus from infected liver cells.

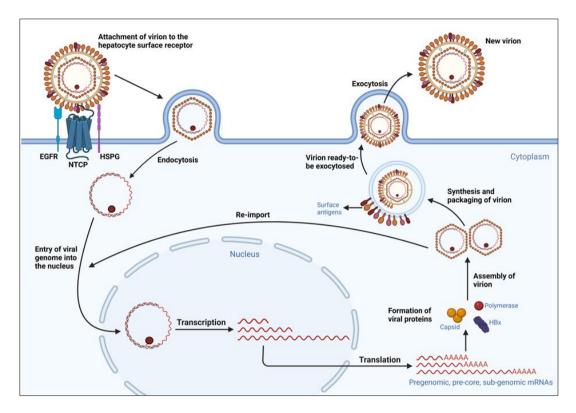


Figure 2 Schematic representation of Hepatitis B viral pathogenesis/ Hepatitis B Virus Pathogenesis: A Schematic Overview

1.1.2. Understanding the progression of Hepatocellular carcinoma by Hepatitis B virus

Hepatitis B virus (HBV) infects the liver and can range from being inactive to a more or less progressive form of hepatitis that can lead to cirrhosis and liver cancer (hepatocellular carcinoma, or HCC) [14, 15, 16]. There are two types of chronic hepatitis B: Hepatitis B envelope Antigen (HBeAg)-positive, which is due to wild-type HBV and represents the early phase of chronic HBV infection, and HBeAg-negative, which is caused by a naturally occurring HBV variant with mutations in certain regions of the genome and represents a late phase of the infection [17]. HBeAg-negative chronic hepatitis B is generally associated with more severe liver disease and a lower response rate to antiviral therapy [17, 18, 19]. The risk of developing cirrhosis within 5 years of being diagnosed with chronic hepatitis B ranges from 8-20%, and the risk of developing liver failure, or HCC, is also significant [20]. HCC is one of the most common cancers worldwide, with about 75% of cases being related to chronic HBV infection [21]. The incidence of HCC varies geographically and is higher in people with advanced liver disease. The only way to improve survival after a diagnosis of HCC is through early detection and treatment such as surgical resection, liver transplantation, or percutaneous ablation. Universal vaccination and new therapeutic agents may help prevent the development of cirrhosis and HCC [22].

1.1.3. Current Treatment of Chronic Hepatitis B

There are currently seven drugs available for the treatment of chronic hepatitis B (CHB), including Interferon alpha, Lamivudine, Adefovir, pegylated interferon alpha-2a, Entecavir, Telbivudine, and Tenofovir [14, 15, 23]. These drugs have varying levels of effectiveness in suppressing the hepatitis B virus and improving clinical outcomes, and can be limited by factors such as poor tolerability, the development of resistance, and the presence of co-infections with other viruses (Table1).

Table 1 Overview of Antiviral drugs for treatment	t of Hepatitis B virus (HBV) infection
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Drug	Description	Effectiveness	Resistance profile	Reference
Interferon-∝	Antiviral, antiproliferative, and immunomodulatory effects	Superior to placebo in undetectability of HBV DNA and HBeAg loss	Poor tolerability	[24, 25]
Lamivudine	Oral drug	Poor resistance profile		[26, 27]
Adefovir	Nucleotide analogue	Improved resistance profile compared to lamivudine but not more effective than lamivudine in viral suppression	Better resistance profile than lamivudine	[28]
Entecavir	Potent anti-HBV agent	High rates of undetectable HBV DNA and low HBeAg seroconversion rates	Low rate of resistance in naïve patients, high genetic barrier to resistance in lamivudine- resistance patients	[29, 30]
Telbivudine	Oral drug	High rates of undetachable HBV DNA and low HBeAg seroconversion ratesLow rate of resistance in naïve patients, moderate rate of resistance in lamivudine- resistance patients		[31,32]
Tenofovir	Nucleotide analogue	e High rates of undetachable HBV DNA and low HBeAg seroconversion rates lamivudine- resistance patients		[33]

1.2. Hepatitis C virus

Table 2 Possible Multifunctional Roles of HCV Gene Products

Gene product	Function	Relevance to malignant formation	Reference
Core	May have immunosuppressive activities through interaction with pathways and C1qR.	Yes	[34] [39]
E2	Interferes with interferon actions; Interacts with cell surface marker CD81.	Yes	[40]
NS3	Viral protease; activates various signal transduction pathways	Yes	[35] [41]
NS5A	Implicated in diverse cellular functions including blocking interferon responses.	Yes	[42][43][44] [45][46][47] [48]
P7, NS2, NS4A, NS4B	Not fully defined	Unknown	[49] [50]

Note: The true biological relevance of these observations is not yet established, especially in regards to the development of hepatocellular carcinoma (HCC).

Hepatitis C virus (HCV) is a type of RNA virus that belongs to the *Flaviviridae* family and is classified in the genus *Hepacivirus*. Its genome is approximately 10 kilobases in length and encodes 10 viral gene products that are divided

into structural and non-structural genes (Fig.3). Some of the proposed functions of HCV gene products may be relevant to the development of cancer, such as the core gene product's interaction with pathways related to apoptosis, signal transduction, transcriptional activation, and transformation [34]. The non-structural proteins of HCV may also play a role in sustaining viral persistence and promoting carcinogenesis, such as the NS3 protein's activation of various signal transduction pathways and the NS5A protein's potential role in blocking interferon responses [35, 36, 37, 38]. However, the true biological relevance of these observations is not yet established, especially in regards to the development of hepatocellular carcinoma (HCC) (Table2).

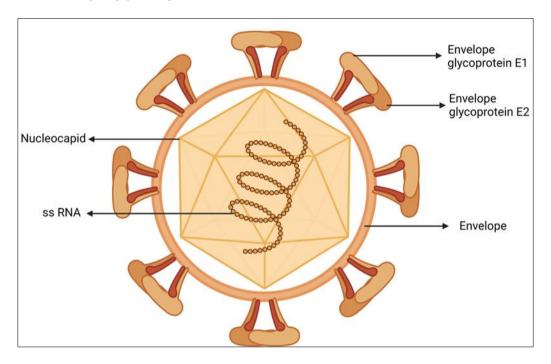
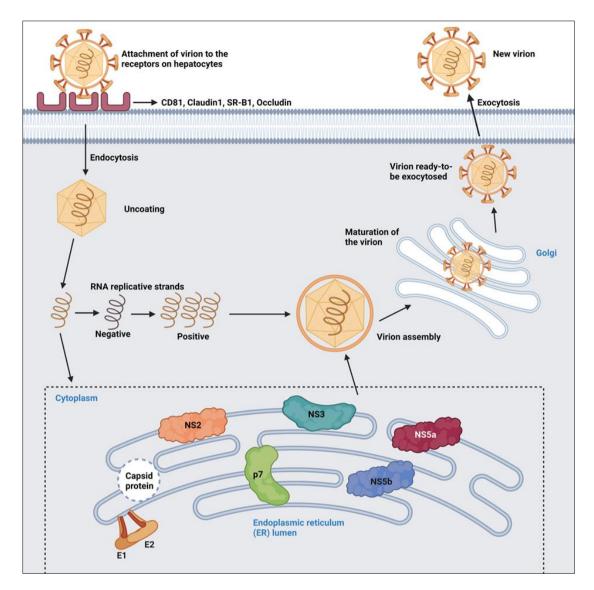
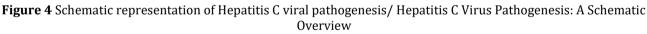


Figure 3 Schematic representation of Hepatitis C

1.2.1. Molecular Mechanisms of HCV pathogenesis

The molecular mechanisms of HCV pathogenesis involve a complex interplay between the virus and the host, with the virus exploiting various host pathways and mechanisms to establish infection and persist in the liver **(Fig.4.).** HCV has been linked to the production of reactive oxygen species (ROS), which can contribute to liver injury and oxidative stress in infected individuals. HCV gene products such as the core and NS5A proteins have been shown to induce ROS production through various mechanisms, including the release of cytochrome C by the core protein and the release of calcium by the NS5A protein [51, 43]. Chronic inflammation caused by HCV infection can also contribute to ROS production. The presence of ROS and the resulting oxidative stress can create a pro-carcinogenic environment that leads to chromosomal damage and increased mutation rates in infected cells. In particular, HCV infection has been associated with mutations in p53, beta-catenin, and other proto-oncogenes and tumour suppressor genes in HCC [52, 53]. A study also found that HCV infection can cause a "hypermutator" phenotype in lymphoma cells, with increased mutational frequencies in certain genes, double-stranded chromosomal breaks, and the activation of error-prone DNA polymerases and activation-induced cytidine deaminase. These findings suggest that hypermutational events may be a mechanism of carcinogenesis during HCV infection[54, 55, 56].





1.2.2. Understanding the Progression of Chronic Hepatitis C Infection

Chronic hepatitis C (HCV) is a viral infection that can cause liver scarring (fibrosis). The progression of fibrosis is a key factor in determining the need for treatment and the overall outlook of the infection [57, 58, 59]. A number of factors, including inflammation and stellate cell activation, contribute to the development of fibrosis. Risk factors for fibrosis progression include age at infection, male gender, heavy alcohol use, and being immunocompromised. Obesity, diabetes, and hepatic steatosis (excess fat accumulation in the liver) may also affect the progression of fibrosis in HCV-infected individuals [60, 61]. There is currently no reliable test to predict the rate of fibrosis progression in a specific case. Normal ALT levels do not necessarily mean a patient is not at risk for fibrosis worsening, but elevated ALT levels are a risk factor[62]. The most accurate way to assess fibrosis progression is to repeat a liver biopsy 3-5 years after the initial biopsy. However, there is a need for improvement and validation in the use of fibrosis serum markers.

1.2.3. Current Treatment of Hepatitis C

Chronic hepatitis C (HCV) is currently treated with a combination of pegylated interferon and ribavirin, which has a success rate of around 50% in achieving sustained virologic response (SVR) [63,64,65,66,67]. However, this treatment has significant side effects and is not well tolerated by many patients. Researchers are currently developing enzyme inhibitors, such as protease and polymerase inhibitors, which show promise in combination with pegylated interferon and ribavirin[68]. It is likely that interferon-based therapy and ribavirin will remain the main treatment for HCV in the coming years, but the use of specifically targeted antiviral therapy drugs could improve SVR rates and eventually lead

to the use of interferon- and ribavirin-sparing regimens[69,70]. Many natural compounds derived from plants have been shown to have antiviral activity and have been studied as potential sources for new drugs. Approximately 200 antiviral agents have been developed in the past 50 years, with around 40% being vaccines and the rest being natural or semi-synthetic compounds inspired by nature. These compounds, including flavonoids, polyphenols, alkaloids, stilbenoids, and terpenes, have been shown to prevent the adhesion, penetration, duplication, or replication of viruses. Some of these compounds have previously been shown to be effective against viruses that are similar to HCV.

Natural products and synthetic therapeutic chemicals can work together to produce potent and effective medicines. This theory is supported by the existing research on the development of new drugs. This strategy has the potential to serve as the foundation for the development of therapeutic natural products that are designed for consumption by humans. Furthermore, the treatment of Hepatitis B and Hepatitis C virus infections, which can lead to liver carcinoma, should consider the use of such plant-based natural products exhibiting elevated antiviral activity. In this context, the current review discusses certain classes of natural compounds that have previously been reported to exhibit antiviral activity against similar viruses.

2. Search methodology

A thorough literature search was conducted for retrieving the studies published until September 2021 in the following databases: PubMed, Science Direct & Google Scholar. Inclusion and exclusion criteria were used to screen out 96 relevant papers. The papers included extensive data on several known and novel natural compounds studied for Hepatitis B & C. **Table 3** provides a concise summary of each of these papers, as well as the mechanism of action of the natural inhibitors that target viral proteins. The papers identified focused on bioactive plant-derived compounds derived from natural sources and their impact on various stages of viral replication, with a preference for compounds with low toxicity and the potential for multi-site inhibition.

3. Natural plant derived inhibitors targeting Hepatitis B and C viral proteins during their replication

In the section that follows, natural products with demonstrated anti-HBV (hepatitis B virus) and anti-HCV (Hepatitis C virus) activities will be discussed, with a particular focus on those that utilize mechanisms that differ from those of currently approved drugs. The aim is to explore the potential of these natural products as alternative or complementary treatments for HBV and HCV infections, and to highlight any unique or novel modes of action that they may possess. Given the significant burden of HBV and HCV on global health, the identification and characterization of these natural products may provide valuable insights for the development of new and more effective treatments for these viral infections.

S.No	Compound	Plant source	Virus	Target protein	Mechanism of action	IC ₅₀ /EC ₅₀	Reference
1.	Rosmarinic acid (phenolic compound)	Lamiaceae	HBV	ε-polymerase	Inhibits replication by binding to ε- polymerase	NR	[73]
2.	Methanolic extract	Hybanthus enneaspermus	HBV	Surface antigen	Entry, replication,		[74,75]
3.	Methanolic extract	Terminalia bellerica	HBV	DNA polymerase	and maturation of HBV particles		
4.	Methanolic extract	Enicostemma axillare	HBV	DNA polymerase	are all inhibited.		
5.	Crude extract	Phyllanthus amarus	HBV	DNA polymerase			
6.	Caudatin	Cynanchum auriculatum	HBV	anti-HBV action by interfering HBV enhancers and promoters	Inhibitory activity against HBsAg secretion and	142.67 μmol/L	[76, 77]

Table 3 List of the natural compounds with anti-Hep B and anti-Hep C activities along with their mechanisms of action.

					HBV DNA replication		
7.	Helioxanthin	Taiwania cryptomerioides	HBV	HBV mRNAs transcription, or in a post- transcriptional manner	To inhibit the HBV RNA and the viral protein expression	0.1 ±0.2mM 32 ±1.4 mM	[78, 79]
8.	Curcumin	Rhizome of <i>Curcuma</i> <i>longa</i> L.	HBV	gluconeogenesi s gene coactivator PGC-1α	Inhibits HBV replication in part by preventing the acetylation of histones bound to cccDNA.	NA	[80, 81, 82]
9.	Asiaticoside	Hydrocotyle sibthorpioides	HBV	core, S1, S2, and X gene promoter activities	Reduced viral DNA transcription and replication	23.5 μmol/L	[83]
10.	Phytoconstitue nts extract	Gymnema sylvestre R.Br.	HBV	Surface Antigen	Inhibits HBsAg and HBV DNA polymerase activity	NR	[84]
11.	LPRP-Et-97543	Liriope platyphylla roots	HBV	Entry of target protein	Controls gene expression and DNA replication	NR	[85]
12.	C- boivinopyranos yl flavones (luteolin-6-C- β - d- boivinopyranos yl- $3'$ - O - β -D- glucopyranosid e and chrysoeriol-6-C- β -D- boivinopyranos yl- $4'$ - O - β -D- glucopyranosid e) extracts	Alternanthera philoxeroides	HBV	Surface Antigen	Reduces the amount of HBsAg that is secreted by HepG2.15	NR	[86]
13.	Dephinidin	Anthocyanidin abundant in Vaccinium corymbosum	HCV	E1 and E2 glyco-proteins	Effects on virus-host interaction via action on E1 and E2 glycoproteins, which results in conformationa l changes in viral particles.	EC ₅₀ - 3.7 + 0.8 μM	[87]
14.	Mangosteen	Garcinia mangostana L	HCV	NS5b	Reduces HCV protein and RNA levels in 1b & 2a	EC ₅₀ – 1b- 5.1 μg/ml	[88]

					infectious replicon systems.	2a- 3.8 μg/ml	
15.	Epigallocatechi n-3-gallate (EGCG)	Camellia sinensis	HCV	E1 and E2 glyco-proteins	EGCG's direct action on E1, E2 envelope glycoprotein alters the viral envelope structure without destroying it and blocks HCV cell binding. Blocks cell-to- cell communicatio	IC ₅₀ - 5-21 μΜ	[89, 90, 91, 92, 93]
16.	Gallic acid	Limonium sinense	HCV	NS's (NS 5A)	n Inhibits entry and replication, inhibits HCV protease function, and downregulates HCV-RNA.		[94, 95]
17.	Apigenin	Petroselinum crispum OR flowers of chamomile Eclipta alba	HCV	NS5B	Has an inhibitory effect on the development and replication of HCV viral particles containing miR122 (microRNA 122). HCV replication can be suppressed by blocking the RNA- dependent RNA polymerase, NS5B, in vitro.	IC ₅₀ - 4.3- 7.9 μM	[96, 97, 100, 101]
18.	Lucidone	Lindera erythrocarpa	HCV	NS3/4A	Increases IFN response and blocks NS3/4A protease by upregulating HO-1.	IC ₅₀ - 1.1 μΜ	[102]
19.	Vitisin B	Vitis vinifera	HCV	NS3	Inhibitor of HCV replication that targets NS3 helicase	IC ₅₀ - 0.006 μM or 6nM	[103]

20.	Saikosaponin	Bupleurum kaoi	HCV	E2	Blocks viral	16.13 +	
	B2				entry by	2.41 μM	[104]
					neutralizing		
					the viral		
					particles		
21.	Honokiol	Magnolia	HCV	Components of	Have multiple	(LD50/	[105]
		officinalis		replication	effects on HCV	EC90 = 5.4	
				complex, NS3,	infection,		
				NS5A and	inhibiting		
				NS5B, were	entry,		
				downregulated	translation		
					and		
					replication		
22.	Naringenin	Grapefruit	HCV	Against NS2	Inhibits HCV	109 µM	[106]
		flavonoid		protease	assembly, as it		
					reduces the		
					buildup of		
					infectious		
					particles		
					within cells.		

3.1. Natural compounds targeting viral proteins associated with Hepatitis B for therapeutic intervention

As summarised in the Table.3. Rosmarinic acid is a natural compound that is found in abundance in various herbs belonging to the *Lamiaceae* family, such as spearmint, sage, peppermint, and perilla. It is commonly used as a dietary supplement and in Chinese herbal medicine. This compound has been shown to inhibit the binding of ε -Pol without affecting the binding of dsRNA-RIG-I, the helicase activity of RIG-I, or the binding of ε -ISG20 [71, 72]. Therefore, it is believed that rosmarinic acid does not interfere with the host's antiviral immune response. In vitro studies have also shown that rosmarinic acid treatment strongly abolishes ε -Pol binding, and it has been demonstrated to suppress HBV replication in cells [73].

Inhibitors of viral entry and fusion are receiving increasing attention for HBV treatment due to the highly selective tropism of the virus. Methanolic extracts of *Hybanthus enneaspermus* have been shown to inhibit HBs Ag binding, while methanolic extracts from seeds of *Terminalia bellerica* and leaves of *Enicostemma axillare* have been demonstrated to block HBV DNA polymerase. *Phyllanthus amarus* extracts have been found to downregulate hepatitis B virus mRNA transcription, suppress hepatitis B virus polymerase activity, and inhibit the release of the virus into Hep-G/2.2.15 cells [74]. The antiviral activity of these three plants was further investigated and it was found that the methanol extract of *Hybanthus enneaspermus* inhibited HBs Ag binding, while methanolic extracts of *Terminalia bellerica* and *Enicostemma axillare* inhibited HBV DNA polymerase. However, none of the three plants exhibited inhibition of both HBs Ag binding and HBV DNA polymerase, indicating that simply screening for antiviral activity using a single assay is not conclusive proof and further molecular studies are necessary. These studies also revealed the HBV receptor binding capability of all three plants. While there are no published antiviral studies on these three plants, there are numerous other plants that have been studied elsewhere and their results are cited for comparison. The methanol extract of *Hybanthus enneaspermus* was found to inhibit both HBs Ag binding and HBV DNA polymerase, while only the methanolic extracts of *Terminalia bellerica* and *Enicostemma axillare* inhibited HBS Ag binding and HBV DNA polymerase.

Caudatin is a steroidal compound found in the plant *Cynanchum auriculatum*. It has been shown to have anti-cancer and antiangiogenic properties, meaning it can inhibit the growth of cancer cells and prevent the formation of new blood vessels. Caudatin has also been found to have inhibitory activity against the secretion of HBsAg (a protein produced by the hepatitis B virus) and the replication of HBV DNA. In particular, the compound 3-O-(3,4,5-trimethoxy) cinnamoyl caudatin has been shown to have a novel mechanism of anti-HBV action by interfering with HBV enhancers and promoters. In laboratory studies, caudatin has been shown to cause cell cycle arrest and induce apoptosis (a type of programmed cell death). The IC50 values for caudatin's inhibitory activity against HBsAg secretion and HBV DNA replication have been measured at 142.67 µmol/L (SI = 1.7) and 40.62 mmol/L (SI = 6.0), respectively [76, 77].

Helioxanthin and its derivative are small molecules that have been found to inhibit HBV RNA and viral protein expression. These compounds have unique structures compared to other anti-HBV compounds and may have unique modes of action. In laboratory studies, the treatment of HepG2.2.15 cells with these compounds resulted in the inhibition of HBV mRNA transcripts, including both 3.5 kb and 2.4/2.1 kb mRNAs. This led to a decrease in the HBV core

protein. The 3.5 kb mRNA plays a key role in the HBV life cycle as it encodes the HBV core protein and DNA polymerase, and serves as the template for minus strand DNA synthesis. These results suggest that helioxanthin and 5-4-2 target multiple steps of the viral life cycle and effectively inhibit HBV replication [78, 79].

Curcumin, a natural compound found in the spice turmeric, has been shown to have antiviral properties against HBV (hepatitis B virus) infection. It is believed to inhibit HBV by down-regulating the expression of certain genes, such as PGC-1 α , and increasing the stability of the p53 protein [80, 81]. In a study, researchers examined the effects of curcumin on cccDNA (circular, covalently closed DNA), a form of the HBV genome found in infected liver cells. They found that curcumin was able to reduce the levels of cccDNA-bound histones and overall levels of cccDNA in HBV-infected cells, suggesting it may be a promising agent for the treatment of HBV. Further research is needed to fully understand how curcumin exerts its antiviral effects [82].

Asiaticoside, a compound isolated from the plant *Hydrocotyle sibthorpioides*, has been shown to effectively suppress the levels of HBsAg/HBeAg (proteins produced by the hepatitis B virus), extracellular HBV DNA, and intracellular cccDNA (a form of the HBV genome) in a dose-dependent manner. It also inhibits viral DNA transcription and replication by inhibiting the activity of certain gene promoters, and reduces replication of the hepatitis B virus (DHBV) without causing any obvious signs of toxicity. These findings suggest that asiaticoside may be a promising agent for the treatment of HBV infection [83].

Gymnema sylvestre R. Br. is a plant that has been shown to have antiviral activity. Its active components, known as phytoconstituents, have been shown to inhibit the binding of HBsAg (a protein produced by the hepatitis B virus) and the activity of HBV DNA polymerase, an enzyme involved in the replication of HBV DNA. These findings suggest that *Gymnema sylvestre* may be a useful agent for the treatment of HBV infection [84].

LPRP-Et-97543 is a compound that was isolated from the roots of the plant *Liriope platyphylla*. It has been shown to inhibit the mode of action of the hepatitis B virus (HBV) by controlling gene expression and DNA replication by viral proteins. This interference with the viral proteins disrupts the nuclear factor NF- κ B pathway, which is a signaling pathway that plays a role in the regulation of immune and inflammatory responses. These findings suggest that LPRP-Et-97543 may be a promising agent for the treatment of HBV infection [85].

Two new compounds called luteolin-6-C- β -d-boivinopyranosyl-3'-O- β -D-glucopyranoside and chrysoeriol-6-C- β -D-boivinopyranosyl-4'-O- β -D-glucopyranoside have been identified in the plant *Alternanthera philoxeroides*. These compounds, known as C-boivinopyranosyl flavones, have been shown to have significant anti-HBV (hepatitis B virus) activity. Specifically, they have been found to reduce the secretion of HBsAg, a protein produced by HBV, in HepG2.15 cells, a type of liver cell line. These findings suggest that these C-boivinopyranosyl flavones may be useful agents for the treatment of HBV infection [86].

3.2. Natural compounds targeting viral proteins associated with Hepatitis C for therapeutic intervention

Delphinidin, a plant pigment found in anthocyanidins, has been shown to be a more effective HCV entry inhibitor. It has been observed to inhibit HCV attachment to the cell surface and is effective in primary hepatocytes. It combats the HCV entry through the use of HCV pseudo particle (HCVpp), which harbor E1E2 envelope glycoproteins of various genotypes, indicating that its inhibitory effects are not limited to a specific genotype. In addition to inhibiting HCVpp entry, delphinidin has also been shown to inhibit HCV cell culture (HCVcc) infections, suggesting that it may interfere with the function of the E1E2 envelope glycoprotein on the viral particle. Overall, delphinidin appears to be a promising new HCV entry inhibitor with potential for use in the treatment of HCV infection [87].

Garcinia mangostana, also known as Mangosteen, is a plant native to Indonesia, Malaysia, the Philippines, and Thailand that has been shown to have antioxidant and antiviral properties. Researchers have hypothesized that it may have therapeutic potential against HCV infection, and a study found that the ethanol extract of mangosteen (MG-EtOH) had the most potent anti-HCV replication activity. Further analysis identified two molecules, α - and γ -mangostins, as the major contributors to this antiviral effect. The study also found that MG-EtOH was able to restore normal levels of ROS production in HCV-infected cells, suggesting that its ROS-scavenging activity may be involved in its inhibitory effect on HCV replication [88].

Epigallocatechin-3-gallate (EGCG), a flavonoid found in green tea, has been shown to inhibit HCV entry. It has been tested in HCVcc and HCVpp systems, as well as in primary human hepatocytes, and has been shown to directly act on the viral particle to prevent attachment to the cell surface [89, 90, 91]. EGCG has also been observed to have a pangenotypic effect against HCV, meaning it is effective against all genotypes of the virus. It is thought that EGCG may alter

the structure of the HCV envelope by acting on the E1E2 envelope glycoprotein, without disrupting it, leading to the blockade of HCV binding to cells. EGCG has also been suggested to inhibit the binding of the HCV envelope to cell surface heparan sulphate [92, 93]. Gallic acid is a type of phenolic acid that has been identified as an anti-HCV (hepatitis C virus) agent when isolated from grape seed extract. It has also been isolated from a plant called Limonium sinense, which belongs to the *Plumbaginaceae* family and is commonly used in traditional medicine. When a root water extract from L. sinense was tested, it was found to inhibit HCV infection at the entry step, which refers to the initial stage of the virus entering and infecting a host cell. This inhibition occurred more specifically during the attachment and fusion/endocytosis processes, which involve the virus attaching to and merging with the host cell membrane. Gallic acid was found to be the most active in inhibiting this process, with an inhibitory activity on viral entry with an IC50 value of 36.4 (µM) [94, 95]. Apigenin is a flavonoid found in certain plants that has been shown to inhibit the replication of hepatitis C virus (HCV) by blocking the maturation of microRNA (miRNA) called miR122 [96, 97]. miR122 is essential for the replication of HCV RNA in liver cells [98, 99]. Research has also shown that apigenin and another flavonoid called luteolin can inhibit HCV infection and replication in cells expressing HCV replicon [100]. These compounds were identified using a pharmacophore and structure-based study targeting NS5B, and the anti-NS5B polymerase activity of luteolin was confirmed in vitro. Apigenin and luteolin extracted from the plant *Eclipta alba*, which is used in Ayurvedic medicine, have also been shown to inhibit HCV replication by inhibiting NS5B RNA-dependent RNA polymerase in vitro and in cells expressing a sub genomic replicon [101]. Lucidone, a compound isolated from the fruit of the plant Lindera erythrocarpa, has been shown to specifically inhibit the replication of hepatitis C virus (HCV) RNA. L. erythrocarpa, a plant native to Asia, has traditionally been used in folk medicine and its fruit has a range of pharmacological properties. Research has shown that HCV RNA levels are suppressed by lucidone in a concentration-dependent manner, with an EC50 (concentration required to achieve 50% effectiveness) of $15 \pm 0.5 \mu$ M in HCV replicon cells. The compound was also found to have a CC50 (concentration required to achieve 50% cytotoxicity) of $620 \pm 5 \,\mu$ M, indicating that it is not cytotoxic at effective antiviral concentrations. An infectious assay confirmed the inhibitory effect of lucidone on viral RNA replication with an EC50 of 20 \pm 1.1 μ M, and a selectivity index (SI; CC50/EC50) of approximately 31, suggesting that it could be a promising lead compound for the development of new anti-HCV agents [102]. The resveratrol tetramer Vitisin B, which is found in the root of grapevines, has been shown to have the highest anti-hepatitis C virus (HCV) replication activity. Further analysis of several HCV variants resistant to vitisin B, as well as in vitro binding and helicase assays, suggests that the mode of action of vitisin B is the inhibition of the viral helicase NS3. Vitisin B was found to have the greatest activity against HCV replication, and it is thought that its direct binding to and inhibition of HCV NS3 helicase may be an important factor in its ability to effectively suppress HCV replication [103]. Inhibition of early hepatitis C virus (HCV) entry has been demonstrated for Saikosaponin Sb2. Virus particles are neutralised, attachment is prevented, and entry and fusion are blocked. It has been shown that SSb2 acts on the HCV E2 protein through analysis of soluble viral glycoproteins. Furthermore, SSb2 has been shown to prevent the binding of serum-derived HCV to hepatoma cells, as well as inhibit infection by multiple genotypic strains of HCV. Researchers have discovered that SSb2 can prevent HCV infection in primary human hepatocytes when used as a treatment [104]. Honokiol is a natural compound found in the Magnolia officinalis plant that has been shown to have several pharmacological effects, including anti-inflammatory and anti-cancer properties. In pre-clinical studies, honokiol has demonstrated effectiveness in inhibiting the replication of the hepatitis C virus (HCV) by reducing the expression of proteins that are necessary for HCV infection. This antiviral activity is observed at low concentrations of honokiol and does not appear to be toxic to cells. Honokiol may also inhibit HCV replication by modulating signaling pathways related to reactive oxygen species (ROS), PI3K/Akt, NFκB, and STAT3. When combined with a low dose of interferon-a, honokiol has been shown to have an even more potent inhibitory effect on HCV replication compared to the standard treatment with ribavirin [105]. Naringenin is a flavonoid that is found in grapefruit and is commonly used as a dietary supplement. It has been shown to have anti-oxidant, anti-inflammatory, and anti-carcinogenic properties both in laboratory and animal studies. Naringenin has been found to inhibit the secretion of ApoB and HCV particles in a dose-dependent manner, without affecting the levels of HCV RNA or protein within cells. This suggests that naringenin may prevent the accumulation of infectious particles by blocking the assembly of HCV [106].

4. Conclusion

HBV & HCV, are a major cause of liver cancer and pose a significant threat to global health. As there are only a few drugs against HBV & HCV and no vaccine for HCV, there is an urgent need for the discovery of new and natural agents, having lesser side effects compared to chemical drugs. In this context, several attempts are made to successfully identify the inhibitors of hepatotropic viruses, including peptides, vaccines, small molecule compounds, and even natural products exhibiting anti-viral activity. The present review is, therefore, an attempt to review the existing literature for potential natural inhibitors against hepatotropic viruses causing liver cancer to provide an overview that could assist in further investigations related to this topic of concern. These natural inhibitors are suitable for managing Liver cancer infections through the modulation of a wide range of molecular targets through effective mechanisms of action (Table.3) and minimum toxicity. For example, Rosmarinic acid can be used against HBV which hinders viral replication and

Epigallocatechin-3-gallate (EGCG) alters HCV envelop protein's structure hampering the viral entry in the host. Overall, the data collected from various sources indicated the availability of different classes of compounds with high favourable efficacy are included are flavonoids, flavanones, flavanols, alkaloids, polyphenols, and terpenes. As a future scope, it would be valuable to investigate the use of combinations of these compounds for checking the potential of improvement in overall therapeutic success. Considering the promising and powerful effects of these natural products, they should be further researched, developed and investigated as alternative therapies to current standard treatments.

Compliance with ethical standards

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Disclosure of conflict of interest

The authors declare that there is no conflict of interest with respect to the current study.

References

- [1] Liu Z, Hou J. Hepatitis B virus (HBV) and hepatitis C virus (HCV) dual infection. International Journal of Medical Sciences. 2006, 3(2):57.
- [2] Lamontagne RJ, Bagga S, Bouchard MJ. Hepatitis B virus molecular biology and pathogenesis. Hepatoma research. 2016, 2:163.
- [3] Glebe D, Urban S. Viral and cellular determinants involved in hepadnaviral entry. World journal of gastroenterology: WJG. 2007 Jan 1, 13(1):22.
- [4] Hu J, Liu K. Complete and incomplete hepatitis B virus particles: formation, function, and application. Viruses. 2017 Mar 21, 9(3):56.
- [5] Patient R, Hourioux C, Roingeard P. Morphogenesis of hepatitis B virus and its subviral envelope particles. Cellular microbiology. 2009 Nov, 11(11):1561-70.
- [6] Gavilanes F, Gonzalez-Ros JM, Peterson DL. Structure of hepatitis B surface antigen. Characterization of the lipid components and their association with the viral proteins. Journal of biological chemistry. 1982 Jul 10, 257(13):7770-7.
- [7] Bruss V. Hepatitis B virus morphogenesis. World journal of gastroenterology: WJG. 2007 Jan 1, 13(1):65.
- [8] Zhu YZ, Qian XJ, Zhao P, Qi ZT. How hepatitis C virus invades hepatocytes: the mystery of viral entry. World journal of gastroenterology: WJG. 2014 Apr 7, 20(13):3457.
- [9] Iwamoto M, Saso W, Sugiyama R, Ishii K, Ohki M, Nagamori S, Suzuki R, Aizaki H, Ryo A, Yun JH, Park SY. Epidermal growth factor receptor is a host-entry cofactor triggering hepatitis B virus internalization. Proceedings of the National Academy of Sciences. 2019 Apr 23, 116(17):8487-92.
- [10] Macovei A, Radulescu C, Lazar C, Petrescu S, Durantel D, Dwek RA, Zitzmann N, Nichita NB. Hepatitis B virus requires intact caveolin-1 function for productive infection in HepaRG cells. Journal of virology. 2010 Jan 1, 84(1):243-53.
- [11] Huang HC, Chen CC, Chang WC, Tao MH, Huang C. Entry of hepatitis B virus into immortalized human primary hepatocytes by clathrin-dependent endocytosis. Journal of virology. 2012 Sep 1, 86(17):9443-53.
- [12] Umetsu T, Inoue J, Kogure T, Kakazu E, Ninomiya M, Iwata T, Takai S, Nakamura T, Sano A, Shimosegawa T. Inhibitory effect of silibinin on hepatitis B virus entry. Biochemistry and biophysics reports. 2018 Jul 1, 14:20-5.
- [13] Herrscher C, Pastor F, Burlaud-Gaillard J, Dumans A, Seigneuret F, Moreau A, Patient R, Eymieux S, de Rocquigny H, Hourioux C, Roingeard P. Hepatitis B virus entry into HepG2-NTCP cells requires clathrin-mediated endocytosis. Cellular microbiology. 2020 Aug, 22(8):e13205.
- [14] Hoofnagle JH, Doo E, Liang TJ, Fleischer R, Lok AS. Management of hepatitis B: summary of a clinical research workshop. Hepatology. 2007 Apr, 45(4):1056-75.
- [15] Lok AS, McMahon BJ. Chronic hepatitis B. HEPATOLOGY-BALTIMORE THEN ORLANDO-. 2007 Feb 1, 45(2):507.

- [16] Martinot-Peignoux M, Boyer N, Colombat M, Akremi R, Pham BN, Ollivier S, Castelnau C, Valla D, Degott C, Marcellin P. Serum hepatitis B virus DNA levels and liver histology in inactive HBsAg carriers. Journal of hepatology. 2002 Apr 1, 36(4):543-6.
- [17] Hadziyannis SJ, Vassilopoulos D. Hepatitis B e antigen—negative chronic hepatitis B. Hepatology. 2001 Oct 1, 34(4):617-24.
- [18] Zarski JP, Marcellin P, Leroy V, Trepo C, Samuel D, Ganne-Carrie N, Barange K, Canva V, Doffoel M, Cales P, Fédération Nationale des Pôles de Référence et des Réseaux Hépatites. Characteristics of patients with chronic hepatitis B in France: predominant frequency of HBe antigen negative cases. Journal of hepatology. 2006 Sep 1, 45(3):355-60.
- [19] Fattovich G, Bortolotti F, Donato F. Natural history of chronic hepatitis B: special emphasis on disease progression and prognostic factors. Journal of hepatology. 2008 Feb 1, 48(2):335-52.
- [20] Fattovich G, Stroffolini T, Zagni I, Donato F. Hepatocellular carcinoma in cirrhosis: incidence and risk factors. Gastroenterology. 2004 Nov 1, 127(5):S35-50.
- [21] Bruix J, Sherman M, Llovet JM, Beaugrand M, Lencioni R, Burroughs AK, Christensen E, Pagliaro L, Colombo M, Rodés J. Clinical management of hepatocellular carcinoma. Conclusions of the Barcelona-2000 EASL conference. Journal of hepatology. 2001 Sep 1, 35(3):421-30.
- [22] Liaw YF. Prevention and surveillance of hepatitis B virus-related hepatocellular carcinoma. InSeminars in liver disease 2005 Feb (Vol. 25, No. S 1, pp. 40-47). Copyright© 2005 by Thieme Medical Publishers, Inc., 333 Seventh Avenue, New York, NY 10001, USA.
- [23] European Association For The Study Of The Liver. EASL Clinical Practice Guidelines: management of chronic hepatitis B. Journal of hepatology. 2009 Feb 1, 50(2):227-42.
- [24] Wong DK, Cheung AM, O'Rourke K, Naylor CD, Detsky AS, Heathcote J. Effect of alpha-interferon treatment in patients with hepatitis B e antigen-positive chronic hepatitis B: a meta-analysis. Annals of internal medicine. 1993 Aug 15, 119(4):312-23.
- [25] Niederau C, Heintges T, Lange S, Goldmann G, Niederau CM, Mohr L, Häussinger D. Long-term follow-up of HBeAgpositive patients treated with interferon alfa for chronic hepatitis B. New England Journal of Medicine. 1996 May 30, 334(22):1422-7.
- [26] Liaw YF, Leung NW, Chang TT, Guan R, Tai DI, Ng KY, Chien RN, Dent J, Roman L, Edmundson S, Lai CL. Effects of extended lamivudine therapy in Asian patients with chronic hepatitis B. Gastroenterology. 2000 Jul 1, 119(1):172-80.
- [27] Zoulim F, Perrillo R. Hepatitis B: reflections on the current approach to antiviral therapy. Journal of hepatology. 2008 Jan 1, 48:S2-19.
- [28] Hadziyannis SJ, Tassopoulos NC, Heathcote EJ, Chang TT, Kitis G, Rizzetto M, Marcellin P, Lim SG, Goodman Z, Ma J, Brosgart CL. Long-term therapy with adefovir dipivoxil for HBeAg-negative chronic hepatitis B for up to 5 years. Gastroenterology. 2006 Dec 1, 131(6):1743-51.
- [29] Colonno RJ. Four year assessment of ETV resistance in nucleosidenaive and lamivudine refractory patients. J Hepatol.. 2007, 46:S294.
- [30] Sherman M, Yurdaydin C, Sollano J, Silva M, Liaw YF, Cianciara J, Boron-Kaczmarska A, Martin P, Goodman Z, Colonno R, Cross A. Entecavir for treatment of lamivudine-refractory, HBeAg-positive chronic hepatitis B. Gastroenterology. 2006 Jun 1, 130(7):2039-49.
- [31] Lai CL, Gane E, Liaw YF, Hsu CW, Thongsawat S, Wang Y, Chen Y, Heathcote EJ, Rasenack J, Bzowej N, Naoumov NV. Telbivudine versus lamivudine in patients with chronic hepatitis B. New England Journal of Medicine. 2007 Dec 20, 357(25):2576-88.
- [32] Lai CL, Gane E, Hsu CW, Thongsawat S, Wang YM, Chen YG, Heathcote EJ, Rasenack J, Bzowej N, Naoumov N, Zeuzem S. Two-year results from the GLOBE trial in patients with hepatitis B: greater clinical and antiviral efficacy for telbivudine (LdT) vs lamivudine. Hepatology. 2006.
- [33] Marcellin P. Hepatitis B and hepatitis C in 2009. Liver International. 2009 Jan, 29:1-8.
- [34] Lai MM, Ware CF. Hepatitis C virus core protein: possible roles in viral pathogenesis. The hepatitis C viruses. 2000:117-34.

- [35] Sakamuro D, Furukawa T, Takegami T. Hepatitis C virus nonstructural protein NS3 transforms NIH 3T3 cells. Journal of virology. 1995 Jun, 69(6):3893-6.
- [36] Ray RB, Lagging LM, Meyer K, Ray R. Hepatitis C virus core protein cooperates with ras and transforms primary rat embryo fibroblasts to tumorigenic phenotype. Journal of Virology. 1996 Jul, 70(7):4438-43.
- [37] Park JS, Yang JM, Min MK. Hepatitis C virus nonstructural protein NS4B transforms NIH3T3 cells in cooperation with the Ha-ras oncogene. Biochemical and biophysical research communications. 2000 Jan 19, 267(2):581-7.
- [38] Gale Jr M, Kwieciszewski B, Dossett M, Nakao H, Katze MG. Antiapoptotic and oncogenic potentials of hepatitis C virus are linked to interferon resistance by viral repression of the PKR protein kinase. Journal of Virology. 1999 Aug 1, 73(8):6506-16.
- [39] Kittlesen DJ, Chianese-Bullock KA, Yao ZQ, Braciale TJ, Hahn YS. Interaction between complement receptor gC1qR and hepatitis C virus core protein inhibits T-lymphocyte proliferation. The Journal of clinical investigation. 2000 Nov 15, 106(10):1239-49.
- [40] Tseng CT, Klimpel GR. Binding of the hepatitis C virus envelope protein E2 to CD81 inhibits natural killer cell functions. The Journal of experimental medicine. 2002 Jan 7, 195(1):43-50.
- [41] Borowski P, Heiland M, Feucht H, Laufs R. Characterisation of non-structural protein 3 of hepatitis C virus as modulator of protein phosphorylation mediated by PKA and PKC: evidences for action on the level of substrate and enzyme. Archives of virology. 1999 Apr, 144(4):687-701.
- [42] Gale Jr MJ, Korth MJ, Tang NM, Tan SL, Hopkins DA, Dever TE, Polyak SJ, Gretch DR, Katze MG. Evidence that hepatitis C virus resistance to interferon is mediated through repression of the PKR protein kinase by the nonstructural 5A protein. Virology. 1997 Apr 14, 230(2):217-27.
- [43] Gong G, Waris G, Tanveer R, Siddiqui A. Human hepatitis C virus NS5A protein alters intracellular calcium levels, induces oxidative stress, and activates STAT-3 and NF-κB. Proceedings of the National Academy of Sciences. 2001 Aug 14, 98(17):9599-604.
- [44] Bain C, Fatmi A, Zoulim F, Zarski JP, Trépo C, Inchauspé G. Impaired allostimulatory function of dendritic cells in chronic hepatitis C infection. Gastroenterology. 2001 Feb 1, 120(2):512-24.
- [45] Khabar KS, Polyak SJ. Hepatitis C virus-host interactions: the NS5A protein and the interferon/chemokine systems. Journal of Interferon & Cytokine Research. 2002 Oct 1, 22(10):1005-12.
- [46] He Y, Nakao H, Tan SL, Polyak SJ, Neddermann P, Vijaysri S, Jacobs BL, Katze MG. Subversion of cell signaling pathways by hepatitis C virus nonstructural 5A protein via interaction with Grb2 and P85 phosphatidylinositol 3-kinase. Journal of virology. 2002 Sep 15, 76(18):9207-17.
- [47] Chung KM, Song OK, Jang SK. Hepatitis C virus nonstructural protein 5A contains potential transcriptional activator domains. Molecules & Cells (Springer Science & Business Media BV). 1997 Oct 31, 7(5).
- [48] Lan KH, Sheu ML, Hwang SJ, Yen SH, Chen SY, Wu JC, Wang YJ, Kato N, Omata M, Chang FY, Lee SD. HCV NS5A interacts with p53 and inhibits p53-mediated apoptosis. Oncogene. 2002 Jul, 21(31):4801-11.
- [49] Lundin M, Monné M, Widell A, Von Heijne G, Persson MA. Topology of the membrane-associated hepatitis C virus protein NS4B. Journal of virology. 2003 May 1, 77(9):5428-38.
- [50] Gao L, Aizaki H, He JW, Lai MM. Interactions between viral nonstructural proteins and host protein hVAP-33 mediate the formation of hepatitis C virus RNA replication complex on lipid raft. Journal of virology. 2004 Apr 1, 78(7):3480-8.
- [51] Okuda M, Li K, Beard MR, Showalter LA, Scholle F, Lemon SM, Weinman SA. Mitochondrial injury, oxidative stress, and antioxidant gene expression are induced by hepatitis C virus core protein. Gastroenterology. 2002 Feb 1, 122(2):366-75.
- [52] Kageyama F, Kobayashi Y, Kawasaki T, Toyokuni S, Uchida K, Nakamura H. Successful interferon therapy reverses enhanced hepatic iron accumulation and lipid peroxidation in chronic hepatitis C. The American journal of gastroenterology. 2000 Apr 1, 95(4):1041-50.
- [53] Huang H, Fujii H, Sankila A, Mahler-Araujo BM, Matsuda M, Cathomas G, Ohgaki H. β-Catenin mutations are frequent in human hepatocellular carcinomas associated with hepatitis C virus infection. The American journal of pathology. 1999 Dec 1, 155(6):1795-801.

- [54] Kitay-Cohen Y, Amiel A, Hilzenrat N, Buskila D, Ashur Y, Fejgin M, Gaber E, Safadi R, Tur-kaspa R, Lishner M. Bcl-2 rearrangement in patients with chronic hepatitis C associated with essential mixed cryoglobulinemia type II. Blood, The Journal of the American Society of Hematology. 2000 Oct 15, 96(8):2910-2.
- [55] Zignego AL, Ferri C, Giannelli F, Giannini C, Caini P, Monti M, Marrocchi ME, Di Pietro E, La Villa G, Laffi G, Gentilini P. Prevalence of bcl-2 rearrangement in patients with hepatitis C virus-related mixed cryoglobulinemia with or without B-cell lymphomas. Annals of internal medicine. 2002 Oct 1, 137(7):571-80.
- [56] Machida K, Cheng KT, Sung VM, Shimodaira S, Lindsay KL, Levine AM, Lai MY, Lai MM. Hepatitis C virus induces a mutator phenotype: enhanced mutations of immunoglobulin and protooncogenes. Proceedings of the National Academy of Sciences. 2004 Mar 23, 101(12):4262-7.
- [57] Marcellin P, Asselah T, Boyer N. Fibrosis and disease progression in hepatitis C. Hepatology. 2002 Nov, 36(S1):S47-56.
- [58] Hoofnagle JH. Course and outcome of hepatitis C. Hepatology. 2002 Nov, 36(S1):S21-9.
- [59] Poynard T, Bedossa P, Opolon P. Natural history of liver fibrosis progression in patients with chronic hepatitis C. The Lancet. 1997 Mar 22, 349(9055):825-32.
- [60] Di Martino V, Rufat P, Boyer N, Renard P, Matheron S, Le Moing V, Degott C, Valla D, Marcellin P. The influence of human immunodeficiency virus coinfection on chronic hepatitis C in injection drug users: a long-term retrospective cohort study. Hepatology. 2001 Dec 1, 34(6):1193-9.
- [61] Moucari R, Asselah T, Cazals–Hatem D, Voitot H, Boyer N, Ripault MP, Sobesky R, Martinot–Peignoux M, Maylin S, Nicolas–Chanoine MH, Paradis V. Insulin resistance in chronic hepatitis C: association with genotypes 1 and 4, serum HCV RNA level, and liver fibrosis. Gastroenterology. 2008 Feb 1, 134(2):416-23.
- [62] Martinot-Peignoux M, Boyer N, Cazals-Hatem D, Pham BN, Gervais A, Breton VL, Levy S, Degott C, Valla DC, Marcellin P. Prospective study on anti-hepatitis C virus-positive patients with persistently normal serum alanine transaminase with or without detectable serum hepatitis C virus RNA. Hepatology. 2001 Nov, 34(5):1000-5.
- [63] Sarrazin C, Kieffer TL, Bartels D, Hanzelka B, Müh U, Welker M, Wincheringer D, Zhou Y, Chu HM, Lin C, Weegink C. Dynamic hepatitis C virus genotypic and phenotypic changes in patients treated with the protease inhibitor telaprevir. Gastroenterology. 2007 May 1, 132(5):1767-77.
- [64] Forestier N, Reesink HW, Weegink CJ, McNair L, Kieffer TL, Chu HM, Purdy S, Jansen PL, Zeuzem S. Antiviral activity of telaprevir (VX-950) and peginterferon alfa-2a in patients with hepatitis C. Hepatology. 2007 Sep, 46(3):640-8.
- [65] Zeuzem S. Telaprevir in combination with peginterferon-alpha-2a with or without ribavirin in the treatment of chronic hepatitis C: final results of the PROVE2 study. Hepatology. 2008, 48(4):418A-9A.
- [66] Kwo P. Boceprevir plus peginterferon alfa-2b/ribavirin for treatment of genotype 1 chronic hepatitis C in previously untreated patients: interim results from the HCV SPRINT-1 study. Hepatology. 2008, 48(4):1027A.
- [67] Roberts SK, Cooksley G, Dore GJ, Robson R, Shaw D, Berns H, Hill G, Klumpp K, Najera I, Washington C. Robust antiviral activity of R1626, a novel nucleoside analog: a randomized, placebo-controlled study in patients with chronic hepatitis C. Hepatology. 2008 Aug, 48(2):398-406.
- [68] Pockros PJ, Nelson D, Godofsky E, Rodriguez-Torres M, Everson GT, Fried MW, Ghalib R, Harrison S, Nyberg L, Shiffman ML, Najera I. R1626 plus peginterferon Alfa-2a provides potent suppression of hepatitis C virus RNA and significant antiviral synergy in combination with ribavirin. Hepatology. 2008 Aug, 48(2):385-97.
- [69] Asselah T, Bieche I, Narguet S, Sabbagh A, Laurendeau I, Ripault MP, Boyer N, Martinot-Peignoux M, Valla D, Vidaud M, Marcellin P. Liver gene expression signature to predict response to pegylated interferon plus ribavirin combination therapy in patients with chronic hepatitis C. Gut. 2008 Apr 1, 57(4):516-24.
- [70] Sher A. Antimicrobial activity of natural products from medicinal plants. Gomal Journal of Medical Sciences. 2009, 7(1).
- [71] Sato S, Li K, Kameyama T, Hayashi T, Ishida Y, Murakami S, Watanabe T, Iijima S, Sakurai Y, Watashi K, Tsutsumi S. The RNA sensor RIG-I dually functions as an innate sensor and direct antiviral factor for hepatitis B virus. Immunity. 2015 Jan 20, 42(1):123-32.
- [72] Liu Y, Nie H, Mao R, Mitra B, Cai D, Yan R, Guo JT, Block TM, Mechti N, Guo H. Interferon-inducible ribonuclease ISG20 inhibits hepatitis B virus replication through directly binding to the epsilon stem-loop structure of viral RNA. PLoS pathogens. 2017 Apr 11, 13(4):e1006296.

- [73] Tsukamoto Y, Ikeda S, Uwai K, Taguchi R, Chayama K, Sakaguchi T, Narita R, Yao WL, Takeuchi F, Otakaki Y, Watashi K. Rosmarinic acid is a novel inhibitor for Hepatitis B virus replication targeting viral epsilon RNA-polymerase interaction. PLoS One. 2018 May 21, 13(5):e0197664.
- [74] Musarra-Pizzo M, Pennisi R, Ben-Amor I, Mandalari G, Sciortino MT. Antiviral activity exerted by natural products against human viruses. Viruses. 2021 May, 13(5):828.
- [75] Al-Jebouri MM, Madish SA. WORLD JOURNAL OF PHARMACY AND PHARMACEUTICAL SCIENCES.
- [76] Wang LJ, Geng CA, Ma YB, Luo J, Huang XY, Chen H, Zhou NJ, Zhang XM, Chen JJ. Design, synthesis, and molecular hybrids of caudatin and cinnamic acids as novel anti-hepatitis B virus agents. European journal of medicinal chemistry. 2012 Aug 1, 54:352-65.
- [77] Wu YH. Naturally derived anti-hepatitis B virus agents and their mechanism of action. World journal of gastroenterology. 2016 Jan 1, 22(1):188.
- [78] Kwon JA, Rho HM. Hepatitis B viral core protein activates the hepatitis B viral enhancer II/pregenomic promoter through the nuclear factor κB binding site. Biochemistry and cell biology. 2002 Aug 1, 80(4):445-55.
- [79] Li Y, Fu L, Yeo H, Zhu JL, Chou CK, Kou YH, Yeh SF, Gullen E, Austin D, Cheng YC. Inhibition of hepatitis B virus gene expression and replication by helioxanthin and its derivative. Antiviral Chemistry and Chemotherapy. 2005 Jun, 16(3):193-201.
- [80] Rechtman MM, Har-Noy O, Bar-Yishay I, Fishman S, Adamovich Y, Shaul Y, Halpern Z, Shlomai A. Curcumin inhibits hepatitis B virus via down-regulation of the metabolic coactivator PGC-1α. FEBS letters. 2010 Jun 3, 584(11):2485-90.
- [81] Kim HJ, Yoo HS, Kim JC, Park CS, Choi MS, Kim M, Choi H, Min JS, Kim YS, Yoon SW, Ahn JK. Antiviral effect of Curcuma longa Linn extract against hepatitis B virus replication. Journal of ethnopharmacology. 2009 Jul 15, 124(2):189-96.
- [82] Wei ZQ, Zhang YH, Ke CZ, Chen HX, Ren P, He YL, Hu P, Ma DQ, Luo J, Meng ZJ. Curcumin inhibits hepatitis B virus infection by down-regulating cccDNA-bound histone acetylation. World Journal of Gastroenterology. 2017 Sep 9, 23(34):6252.
- [83] Huang Q, Zhang S, Huang R, Wei L, Chen Y, Lv S, Liang C, Tan S, Zhuo L, Lin X. Isolation and identification of an anti-hepatitis B virus compound from Hydrocotyle sibthorpioides Lam. Journal of ethnopharmacology. 2013 Nov 25, 150(2):568-75.
- [84] Subashini MS, Rajendran P. In vitro screening of anti HBV and anti HIV properties of Gymnema sylvestre R. Br leaves from Kolli Hills, Tamilnadu, India. Int J Curr Microbiol Appl Sci. 2015, 4:542-7.
- [85] Huang TJ, Tsai YC, Chiang SY, Wang GJ, Kuo YC, Chang YC, Wu YY, Wu YC. Anti-viral effect of a compound isolated from Liriope platyphylla against hepatitis B virus in vitro. Virus Research. 2014 Nov 4, 192:16-24.
- [86] Li B, Guo QL, Tian Y, Liu SJ, Wang Q, Chen L, Dong JX. New anti-HBV C-boivinopyranosyl flavones from Alternanthera philoxeroides. Molecules. 2016 Mar 14, 21(3):336.
- [87] Calland N, Sahuc ME, Belouzard S, Pène V, Bonnafous P, Mesalam AA, Deloison G, Descamps V, Sahpaz S, Wychowski C, Lambert O. Polyphenols inhibit hepatitis C virus entry by a new mechanism of action. Journal of virology. 2015 Oct 1, 89(19):10053-63.
- [88] Choi M, Kim YM, Lee S, Chin YW, Lee C. Mangosteen xanthones suppress hepatitis C virus genome replication. Virus genes. 2014 Oct, 49(2):208-22.
- [89] Calland N, Albecka A, Belouzard S, Wychowski C, Duverlie G, Descamps V, Hober D, Dubuisson J, Rouillé Y, Séron K. (-)-Epigallocatechin-3-gallate is a new inhibitor of hepatitis C virus entry. Hepatology. 2012 Mar, 55(3):720-9.
- [90] Calland N, Sahuc ME, Belouzard S, Pène V, Bonnafous P, Mesalam AA, Deloison G, Descamps V, Sahpaz S, Wychowski C, Lambert O. Polyphenols inhibit hepatitis C virus entry by a new mechanism of action. Journal of virology. 2015 Oct 1, 89(19):10053-63.
- [91] Ciesek S, von Hahn T, Colpitts CC, Schang LM, Friesland M, Steinmann J, Manns MP, Ott M, Wedemeyer H, Meuleman P, Pietschmann T. The green tea polyphenol, epigallocatechin-3-gallate, inhibits hepatitis C virus entry. Hepatology. 2011 Dec, 54(6):1947-55.

- [92] Fukazawa H, Suzuki T, Wakita T, Murakami Y. A cell-based, microplate colorimetric screen identifies 7, 8benzoflavone and green tea gallate catechins as inhibitors of the hepatitis C virus. Biological and Pharmaceutical Bulletin. 2012 Aug 1, 35(8):1320-7.
- [93] Colpitts CC, Schang LM. A small molecule inhibits virion attachment to heparan sulfate-or sialic acid-containing glycans. Journal of virology. 2014 Jul 15, 88(14):7806-17.
- [94] Sharaf M, El-Deeb NM, EL-Adawi HI. The potentiality of grape seed extract as a novel anti-hepatitis C virus agent. Journal of Medical Sciences. 2012 May 20, 12(4):107.
- [95] Hsu WC, Chang SP, Lin LC, Li CL, Richardson CD, Lin CC, Lin LT. Limonium sinense and gallic acid suppress hepatitis C virus infection by blocking early viral entry. Antiviral research. 2015 Jun 1, 118:139-47.
- [96] Ohno M, Shibata C, Kishikawa T, Yoshikawa T, Takata A, Kojima K, Akanuma M, Kang YJ, Yoshida H, Otsuka M, Koike K. The flavonoid apigenin improves glucose tolerance through inhibition of microRNA maturation in miRNA103 transgenic mice. Scientific reports. 2013 Aug 30, 3(1):1-7.
- [97] Shibata C, Ohno M, Otsuka M, Kishikawa T, Goto K, Muroyama R, Kato N, Yoshikawa T, Takata A, Koike K. The flavonoid apigenin inhibits hepatitis C virus replication by decreasing mature microRNA122 levels. Virology. 2014 Aug 1, 462:42-8.
- [98] Gentzsch J, Hinkelmann B, Kaderali L, Irschik H, Jansen R, Sasse F, Frank R, Pietschmann T. Hepatitis C virus complete life cycle screen for identification of small molecules with pro-or antiviral activity. Antiviral research. 2011 Feb 1, 89(2):136-48.
- [99] Randall G, Panis M, Cooper JD, Tellinghuisen TL, Sukhodolets KE, Pfeffer S, Landthaler M, Landgraf P, Kan S, Lindenbach BD, Chien M. Cellular cofactors affecting hepatitis C virus infection and replication. Proceedings of the National Academy of Sciences. 2007 Jul 31, 104(31):12884-9.
- [100] Liu MM, Zhou L, He PL, Zhang YN, Zhou JY, Shen Q, Chen XW, Zuo JP, Li W, Ye DY. Discovery of flavonoid derivatives as anti-HCV agents via pharmacophore search combining molecular docking strategy. European journal of medicinal chemistry. 2012 Jun 1, 52:33-43.
- [101] Manvar D, Mishra M, Kumar S, Pandey VN. Identification and evaluation of anti-hepatitis C virus phytochemicals from Eclipta alba. Journal of ethnopharmacology. 2012 Dec 18, 144(3):545-54.
- [102] Chen WC, Wang SY, Chiu CC, Tseng CK, Lin CK, Wang HC, Lee JC. Lucidone suppresses hepatitis C virus replication by Nrf2-mediated heme oxygenase-1 induction. Antimicrobial agents and chemotherapy. 2013 Mar, 57(3):1180-91.
- [103] Lee S, Yoon KD, Lee M, Cho Y, Choi G, Jang H, Kim B, Jung DH, Oh JG, Kim GW, Oh JW. Identification of a resveratrol tetramer as a potent inhibitor of hepatitis C virus helicase. British journal of pharmacology. 2016 Jan, 173(1):191-211.
- [104] Lin LT, Chung CY, Hsu WC, Chang SP, Hung TC, Shields J, Russell RS, Lin CC, Li CF, Yen MH, Tyrrell DL. Saikosaponin b2 is a naturally occurring terpenoid that efficiently inhibits hepatitis C virus entry. Journal of hepatology. 2015 Mar 1, 62(3):541-8.
- [105] Lan KH, Wang YW, Lee WP, Lan KL, Tseng SH, Hung LR, Yen SH, Lin HC, Lee SD. Multiple effects of Honokiol on the life cycle of hepatitis C virus. Liver International. 2012 Jul, 32(6):989-97.
- [106] Goldwasser J, Cohen PY, Lin W, Kitsberg D, Balaguer P, Polyak SJ, Chung RT, Yarmush ML, Nahmias Y. Naringenin inhibits the assembly and long-term production of infectious hepatitis C virus particles through a PPAR-mediated mechanism. Journal of hepatology. 2011 Nov 1, 55(5):963-71.